Introduction

Congenital heart disease (CHD) represents the most common form of birth defect (1), with a majority of CHDs involving anomalies in cardiac septation. Cardiac septation involves crosstalk between multiple cardiac cell lineages, but mechanisms by which this crosstalk occurs remain far from understood. Mutations in the T-box transcription factor (TF) Tbx20 are associated with a diverse array of CHDs, including dilated cardiomyopathy, double-outlet right ventricle (DORV), and defects in cardiac septation and valvulogenesis (2–6). The diversity of cardiac pathologies associated with Tbx20 mutations underscores the importance of Tbx20 for multiple aspects of heart development.

In mice, previous studies with a hypomorphic mutant of Tbx20 demonstrated a requirement for Tbx20 in outflow tract (OFT) remodeling and endocardial cushion formation (7). Mice that are globally null for Tbx20 are embryonically lethal around E10 and lack specialized chamber myocardium (7–10). Early lethality of the global mutants prevents the study of later roles of Tbx20.

To understand the mechanisms through which mutations in Tbx20 cause CHD, it is critical to elucidate the tissue-specific roles for Tbx20. Recently, Tbx20 was shown to be required in nuclear factor of activated T cells 1–Cre (Nfatc1-Cre) lineages for valve maturation and elongation via regulation of Wnt signaling (11). Here, our analysis of Tie2-Cre Tbx20fl/null mutants and Nfatc1-Cre Tbx20fl/null mutants has revealed broader requirements for Tbx20 in endocardium for multiple aspects of cardiac septation, including normal formation of the OFT and the dorsal mesenchymal protrusion. To understand the molecular and cellular behaviors underlying OFT alignment and septation defects, we combined detailed histological analyses with the generation of several genome-wide data sets from whole heart and purified endocardial lineages, including microarray, Tbx20 ChIP-seq, and ATAC-seq. We intersected these genome-wide data sets with chromatin loop maps to identify long-range–acting, Tbx20-targeted enhancers. Altogether, we believe our data provide new insights into the mechanisms by which mutations in Tbx20 result in cardiac septation defects, highlighting distinct roles for Tbx20 in endocardial lineages. Our work illustrates an integrative approach that links molecular mechanisms of TF-regulatory functions to whole-systems biology, using well-defined cells in relatively small numbers.

Results

Tbx20 is required in endocardial lineages for multiple aspects of cardiac septation. To better understand the roles of Tbx20 in endocardium and endocardium-derived cushion mesenchyme, Tie2-Cre was used to ablate Tbx20. Tie2-Cre Tbx20fl/null mutants were never recovered after birth, indicating prenatal lethality of mutant embryos. Until E13.5, mutant embryos were recovered at expected Mendelian frequencies (Table 1). However, at E14.5 and later, no live mutants were recovered, indicating that Tbx20 is required in endocardial lineages before this stage for embryonic viability.

Mutations in the T-box transcription factor Tbx20 are associated with multiple forms of congenital heart defects, including cardiac septal abnormalities, but our understanding of the contributions of endocardial Tbx20 to heart development remains incomplete. Here, we investigated how Tbx20 interacts with endocardial gene networks to drive the mesenchymal and myocardial movements that are essential for outflow tract and atrioventricular septation. Selective ablation of Tbx20 in murine endocardial lineages reduced the expression of extracellular matrix and cell migration genes that are critical for septation. Using the assay for transposase-accessible chromatin with high-throughput sequencing (ATAC-seq), we identified accessible chromatin within endocardial lineages and intersected these data with Tbx20 ChIP-seq and chromatin loop maps to determine that Tbx20 binds a conserved long-range enhancer to regulate versican (Vcan) expression. We also observed reduced Vcan expression in Tbx20-deficient mice, supporting a direct role for Tbx20 in Vcan regulation. Further, we show that the Vcan enhancer drove reporter gene expression in endocardial lineages in a Tbx20–binding site–dependent manner. This work illuminates gene networks that interact with Tbx20 to orchestrate cardiac septation and provides insight into the chromatin landscape of endocardial lineages during septation.
Table 1. Observed genotype distribution of live embryos recovered from matings between Tie2-Cre Tbx20+/null males and Tbx20fl/fl females

<table>
<thead>
<tr>
<th>Stage</th>
<th>Without Cre</th>
<th>Tbx20fl/fl</th>
<th>Tbx20fl/fl</th>
<th>χ²</th>
</tr>
</thead>
<tbody>
<tr>
<td>E10.5</td>
<td>48 (48.75)</td>
<td>49 (48.75)</td>
<td>48 (48.75)</td>
<td>50 (48.75)</td>
</tr>
<tr>
<td>E12.5</td>
<td>44 (52.5)</td>
<td>44 (52.5)</td>
<td>65 (52.5)</td>
<td>57 (52.5)</td>
</tr>
<tr>
<td>E13.5</td>
<td>22 (25.5)</td>
<td>25 (25.5)</td>
<td>30 (25.5)</td>
<td>25 (25.5)</td>
</tr>
<tr>
<td>E14.5</td>
<td>14 (10)</td>
<td>17 (10)</td>
<td>9 (10)</td>
<td>0 (10)</td>
</tr>
<tr>
<td>E15.5</td>
<td>10 (6.5)</td>
<td>7 (6.5)</td>
<td>9 (6.5)</td>
<td>0 (6.5)</td>
</tr>
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Expected number is in parentheses; χ² results were obtained; *P value of less than 0.05 was considered significant.

At E13.5, mutant embryos displayed overall normal size and morphology (Figure 1A). Histological analysis of mutant hearts at E13.5 revealed completely penetrant cardiac cushion abnormalities and septal defects (Figure 1B). Most notably, mutant hearts had OFT septation defects reminiscent of a DORV, marked by failure of the septal cushion to extend toward the interventricular septum (IVS). OFT and atrioventricular cushions were malformed. Mutant hearts had ventricular septal defects (VSDs) and ostium primum type atrial septal defects (ASDs) (Figure 1C), which are considered a type of atrioventricular septal defect (AVSD) (12). Additionally, the dorsal mesenchymal protrusion (DMP), a second heart field–derived structure that normally appears as a protrusion at the right pulmonary ridge (13), appeared to be lacking in the mutant hearts (Figure 1C). At E10.5 and E13.5, no defects in vascular patterning were observed (Supplemental Figure 1; supplemental material available online with this article; doi:10.1172/JCI85350DS1).

Tbx20 is required for the proliferation of cushion endocardium. To address the question of how a lack of TBX20 might lead to malformed cushions, we analyzed proliferation in cardiac cushion endocardium and mesenchyme at E11.5 using the Rosa26loxCAG-cre lineage indicator. 5-Ethynyl-2′-deoxyuridine (EdU) incorporation rates in endocardium overlying cushions (Tie2-Cre lineage), CD31+; Figure 2A) were decreased by 27% in OFT cushions. A more subtle decrease in atrioventricular cushion endocardium (14%) failed to reach statistical significance (Figure 2B, P < 0.08). The proliferation rates of endocardium-derived mesenchymal cells were unchanged. Accordingly, the overall numbers of cushion mesenchymal cells in E11.5 mutant hearts were similar compared with those in controls (Supplemental Figure 2A). Similar rates of apoptosis were observed in mutant and control cushions (Supplemental Figure 2B).

Tbx20 is required for the normal migratory behavior of endocardial lineages after EMT. Endocardial cells contribute to OFT and atrioventricular cushions by undergoing endothelial-to-mesenchymal transition (EMT) and invading the cardiac jelly (14). Mutant atrioventricular and OFT cushions were colonized by Tie2-Cre Rosa26loxCAG lineage–traced mesenchymal cells (Figure 3A), demonstrating that mutant endocardial cells had undergone EMT. In control OFT cushions, endocardium-derived mesenchymal cells were loosely arranged and intermingled with mesenchymal cells of nonendothelial origin, such as neural crest cells (15). In contrast, mutant cells appeared as dense clusters in the proximal cushions that did not mix with nonendocardium-derived mesenchymal cells and failed to disperse throughout the cushions (Figure 3B). Notably, we also observed these migratory defects by using a more endocardium-restricted Cre line, Nfatc1-Cre (16), to ablate Tbx20 (Supplemental Figure 3, A and B).

To further characterize Tbx20-mutant endocardial cells, we analyzed their ability to undergo EMT in an ex vivo cushion explant assay (17). Mutant OFT cushion endocardial cells underwent EMT to the same extent as did their WT counterparts, as evidenced by abundant numbers of Tie2-Cre lineage–traced cells that migrated away from the explant over the surface of the gel (Figure 3C and Supplemental Figure 3C). In addition, the distances migrated by mutant endocardial cells atop the matrix were comparable to those of control cells (Supplemental Figure 3D). However, there was a striking reduction in the number of mutant endocardium–derived cells able to invade the gel relative to that seen in WT cells (Figure 3, C and D).

Tbx20 in endocardial lineages is required for myocardialization of the OFT. After invasion by endocardial cells and neural crest cells, OFT cushions expand, fuse, and form a mesenchymal outlet septum. Subsequently, cardiomyocytes that flank cushion mesenchyme protrude into cushion mesenchyme. This process, called myocardialization, involves polarized cell movements of OFT cardiomyocytes (18, 19). In control embryos at E13.5, sarcomeric α-actinin–expressing (Actn2-expressing) cardiomyocytes invaded the OFT septum, and cells protruding from the left and right sides met at the base of the proximal OFT septum (Figure 3B). In mutants, however, cardiomyocyte invasion was impaired, with a significant distance separating the protruding cardiomyocytes (Figure 3B). Notably, aberrantly clustered endocardium-derived cells resided in this region in mutants. These observations suggested that mutant endocardial cells were not able to provide the cues to myocytes required for cardiomyocyte invasion.

Tbx20 in endothelial lineages is required for development of the DMP. To better understand the spatiotemporal requirements for Tbx20 in cardiac septation, we used a more endocardium-restricted Cre line, Nfatc1-Cre, to ablate Tbx20. Notably, Nfatc1-Cre Tbx20fl/fl mutants also displayed OFT septation defects, although to a lesser extent than did Tie2-Cre Tbx20fl/fl mutants (Supplemental Figure 3A). However, in contrast to Tie2-Cre Tbx20fl/fl mutants, Nfatc1-Cre Tbx20fl/fl mutants did not show AVSDs at E13.5, and contribution of the DMP to the atrioventricular septal complex (AVD) was evident (Supplemental Figure 3B). Next, we performed genetic lineage tracing to compare Nfatc1-Cre and Tie2-Cre expression domains and found that at E9.5 and E10.5, Nfatc1-Cre did not label pulmonary venous endothelium and did not efficiently label the endocardium lining the sinus venosus and common atrium, whereas these cell populations were robustly labeled by Tie2-Cre (Supplemental Figure 4). Notably, these regions did express Tbx20 (Supplemental Figure 5), suggesting that Tbx20 expression in endothelium adjacent to the DMP, including endothelium of the pulmonary vein, sinus venosus, or atrium, is required for DMP development.

Tbx20 regulates extracellular matrix and cell guidance genes in endocardial lineages. To examine how ablation of Tbx20 affected gene expression, we performed global gene expression analysis on FACS-purified endocardial lineages from E12.5 hearts (Sup-
Genes grouped by the axon guidance term may be of significance to the observed defects in endocardial cell migration as well as myocardialization defects. Axon guidance genes significantly (P < 0.05) downregulated in mutants included roundabout 1 (Robo1) and slit homolog 2 (Slit2), components of the Slit/roundabout signaling pathway that regulate cardiac septation and valve development (24). Myh10, encoding a nonmuscle myosin required for cell motility (25), was also downregulated in Tie2-Cre Tbx20fl/null mutants. Mutations in Myh10 also cause cardiac septation defects (26).

Several additional genes, including the TFs Twist1, Msx1, and Runx1t1, that are associated with cushion development and cardiac septation were also downregulated in Tie2-Cre Tbx20fl/null mutants (27–29). Expression of several genes of potential biological relevance was validated by quantitative real-time PCR (qRT-PCR) (Figure 4B).

Genomic targets of Tbx20 in mid-gestation heart identified by ChIP-seq analysis. TBX20 is a TF that binds DNA and can activate or repress the expression of target genes (30). To identify...
genes directly regulated by TBX20, we mapped TBX20-binding sites in E11.5 mouse heart by ChIP-sequencing (ChIP-seq), using a Tbx20-GFP transgene (31) that drives expression of a TBX20-GFP fusion protein under the control of the Tbx20-regulatory sequences in endocardium, myocardium, and cushion mesenchyme and that closely resembles the endogenous Tbx20 expression pattern seen in embryonic heart (Supplemental Figure 5, C–E). Chromatin extracts prepared from Tbx20-GFP-transgenic hearts were subjected to ChIP to enrich for TBX20-bound chromatin and subjected to massive parallel sequencing. ChIP-seq analysis resulted in 2,988 peaks of TBX20 binding at a 0.1% FDR. Approximately 18% of these peaks were in promoter regions of RefSeq genes, indicating that, although TBX20 directly binds the promoter regions of several genes, the majority of its regulation relies on binding to longer-range DNA elements (Figure 5A).

TF motif discovery (32) within TBX20-bound regions revealed that the majority of regions (56%) had a TBX20 motif (Figure 5B and ref. 31). Canonical T-box motifs (Tbx5, Eomes; AAGTGTCA) were also enriched, though to a lesser extent (Supplemental Table 2). These data indicated that ChIP-seq fragments were highly enriched for genomic regions directly bound by TBX20. In addition, we found overrepresentation of motifs for GATA family TFs MAFA, a member of the bZIP family of TFs that includes API factors MEF2C, homeodomain TFs NKX2-1 and NKX2-5, and TEAD TF-binding motifs (Figure 5B and Supplemental Table 2). NKX2-5, GATA, MEF2, and TEAD TFs have been previously identified as cofactors of TBX20 in cell lines and adult heart, indicating a high level of functional conservation (3, 30, 31, 33, 34).

Genes proximal to TBX20-binding regions were subsequently analyzed for functional annotation using gene ontology (GO) enrichment analysis (35). In embryonic hearts, TBX20 preferentially bound near genes involved in cardiac chamber development, cardiac ventricle development, and cardiac septum morphogenesis (Figure 5C). It is likely that these terms reflect roles of TBX20 in cardiomyocytes, which make up approximately two-thirds of cells within the heart at this stage (data not shown). Genes involved in heart valve formation, EMT, OFT morphogenesis, mesenchyme development, and endocardial cushion formation were also markedly enriched and might reflect roles of TBX20 in endocardial lineages.

Open chromatin in endocardial lineages identified by ATAC-seq. We had used whole-heart chromatin extracts to map TBX20-binding sites, as it would have been technically challenging to isolate sufficient numbers of the endocardial lineage cells required for ChIP-seq experiments. To define subsets of TBX20-binding sites accessible for transcriptional regulation in endocardial lineages, we applied ATAC-seq (36). This method for mapping chromatin accessibility genome wide utilizes transposase, which will preferentially integrate sequencing adapters in regions of open chromatin such as active promoter regions, enhancers, and insulators and can be performed on relatively small amounts of cells. We performed ATAC-seq on FACS-sorted endocardial lineages from E12.5 hearts. Samples were subjected to massive parallel sequencing, and overlapping peaks from replicate samples were merged to identify high-confidence regions of open chromatin. Peaks were highly enriched at promoter regions (Figure 6A).

A first validation of ATAC-seq data revealed a high read count around promoter regions of genes, including Snai1, ET26 transformation–specific (Ets), and Mxs1, expressed in endocardial lineage cells (Figure 6C and refs. 37–39). In contrast, troponin T (Tnnt2) and myosin heavy chain 7 (Myh7), 2 strictly myocardial genes, did not display significant read counts near their promoters. Several cushion-specific enhancers were also marked by open chromatin, confirming that ATAC-seq marks active promoters and enhancers (Figure 6C and refs. 40–42).

The most highly enriched motif in ATAC-seq regions of open chromatin was a CTCF motif (Figure 6B). CTCF is a TF that binds at chromatin domain boundaries, at enhancers and gene promoters, and inside gene bodies, the majority of which will be marked by open chromatin (43, 44). We also found enrichment for TF motifs of the ETS family, a large family of TFs marked by a highly conserved DNA-binding domain (45). ETS TFs are central regulators of endothelial gene expression, and at least 19 ETS factors are expressed in human endothelial cells (46, 47). Other highly overrepresented motifs are GATA and JUN-API motifs (Supplemental Table 2).

Identification of TBX20-binding sites within open chromatin of endocardial lineages. To identify TBX20-binding sites that could be relevant to endocardial lineage development, we selected TBX20 ChIP-seq peaks within regions of open chromatin in endocardial
enhancers might also be shared between endocardium and myocardium. Additionally, we found significant overrepresentation of ETS motifs and AP1 motifs. These have not previously been identified as cofactors of TBX20 and were specifically enriched in the subset of TBX20 peaks that overlapped with endocardial lineage open chromatin (Figure 6E and Supplemental Table 2).

To identify putative direct targets of TBX20 in endocardial lineages, we selected differentially expressed genes that had TBX20-binding sites in a nearby region of open chromatin. We found 44 putative direct endocardial targets downregulated and 72 putative direct endocardial targets overexpressed in Tie2-Cre Tbx20fl/null mutants (Figure 6F). Among the downregulated putative direct target genes that might impact observed mutant phenotypes were Robo1 and Tbx20 itself (Supplemental Table 3).

Figure 3. TBX20 is required for endocardium-derived cell migration. (A) Tie2-Cre lineage–traced mesenchymal cells (tdTom+) failed to migrate properly in mutants. (B) Mutant mesenchymal cells clustered (arrowheads) and failed to colonize the developing valves (arrows). Muscularization of OFT septum by cardiomyocytes (ACTN2+) was delayed (double arrow). (C) Ex vivo cushion explant showing EMT by mutant endocardium-derived cells (tdTom+; phalloidin+); cell migration over the surface of the collagen gel (middle; migration), and cell invasion into the matrix (right; invasion). Shown are representative images of 6 (Tie2-Cre Tbx20fl/null mutant) and 5 (Tie2-Cre control) explants. (D) Decreased matrix invasion in mutant endocardium-derived cells. *P < 0.05, by 2-way ANOVA with Bonferroni’s post test; mean ± SEM; n = 3. Scale bars: 100 μm (A), 20 μm (B), 200 μm (C, left panel), and 20 μm (C, middle and left panels and inset); tdTom, tdTomato; Phall, phalloidin.
Identification of long-range enhancers using publicly available chromatin loop maps. Our understanding of how alterations in the expression of putative direct targets of TBX20 in endocardial lineages might contribute to the observed phenotypes in Tie2-Cre Tbx20fl/null mutants was limited by the fact that their function has not often been explored in endocardium. Additionally, these direct targets were defined by assigning TBX20 ChIP-seq peaks to the nearest genes. While linear proximity to regulatory sites is often used to predict target genes, many examples exist in which enhancers ignore the nearest genes and act on genes further away (48–50).

We next investigated whether the endocardial genetic programs regulated by TBX20 that we uncovered in mouse hearts are also conserved in humans. 3D-ordered chromatin structures connect long-range enhancers to cognate gene promoters. We postulated that chromatin conformation could be used as a guide in assigning TBX20 ChIP-seq peaks within long-range enhancers to their target genes. We used genome-wide interaction maps generated by high-resolution chromatin conformation capture (HiC) of a human fibroblast cell line that has many characteristics of endocardium-derived mesenchyme (51). By projection of chromatin interactions in the human genome onto orthologous conserved regions of the mouse genome and intersection with our TBX20 ChIP-seq peaks within open chromatin of endocardial lineages, as determined by ATAC-seq, we identified 906 transcription start site–bound (TSS-bound) DNA regions with TBX20 peaks and assigned these peaks to 629 orthologous mouse genes. Notably, TBX20 ChIP-seq peaks and ATAC-seq peaks overlap HiC interactions in excess of approximately 2-fold when compared with heterochromatin regions in human fetal heart identified by the Epigenome Roadmap (52). This observation suggests that the human fibroblast HiC data are useful for uncovering functional regions in mouse endocardium (Supplemental Figure 9D). Interestingly, 25% of these peaks were assigned to the most proximal TSS, whereas others were assigned to more distant genes, suggesting that linear proximity is a poor metric for the functional assignment of target genes for distal enhancers (50). With this approach, we found that 27 TBX20 peaks were associated with downregulated genes and 40 TBX20 peaks with upregulated genes (Figure 6G). Among downregulated putative direct targets of TBX20 with potential long-range–acting enhancers in endocardial lineages were \textit{Vcan}, \textit{Myh10}, and \textit{Runx1t1} (Supplemental Table 4). These data suggest that at least some of the most relevant downstream
targets of TBX20 in mouse endocardium are regulated through enhancers conserved between mouse and humans, giving mechanistic insights into the molecular basis for CHD in patients harboring TBX20 mutations.

*Versican expression is diminished in mutant cushions.* Because mutants with decreased VCAN levels have phenotypes strikingly similar to those of Tie2-Cre Tbx20<sup>fl/null</sup> mutants (22, 23), we further investigated Vcan downregulation as a potential direct target that contributes to the observed Tie2-Cre Tbx20<sup>fl/null</sup> phenotypes. qRT-PCR on RNA from purified endothelial lineages with primers specific for transcript variants of Vcan indicated that expression of all 4 variants examined was decreased in mutants (Supplemental Figure 7A). Furthermore, we found that downregulation of Vcan transcripts had occurred by E10.5 in mutant endocardial cells (Supplemental Figure 7B). To investigate whether decreased mRNA levels resulted in decreased protein levels, we performed anti-VCAN immunostaining. In control hearts, VCAN was highly expressed in OFT and atrioventricular cushions, suggesting its secretion by endocardial lineages and, potentially, by mesenchymal cells of nonendothelial origin (Figure 7). In contrast, in Tie2-Cre Tbx20<sup>fl/null</sup> mutants, VCAN was markedly reduced in OFT and atrioventricular cushions. Mutant cushions expressed normal levels of the mesenchymal cell marker platelet-derived growth factor receptor α (PDGFRα) and showed normal levels of ECM proteoglycans (Supplemental Figure 7, D and E). Moreover, VCAN levels surrounding the main bronchi were similar in controls and mutants (Supplemental Figure 7C), confirming that the reduction of VCAN levels in cushions was specific to a loss of TBX20 in endocardial lineages.

*A Vcan enhancer bound by TBX20 in endocardial lineages drives expression in endocardial lineages in vivo.* Because Vcan was significantly downregulated in Tbx20<sup>fl/mutant</sup> endocardial lineages, and since this reduction may contribute to cardiac defects in our mutant, we further investigated the direct regulation of Vcan by TBX20. Remarkably, the HiC data suggested
Using a conditional mouse model and genome-wide analyses, we investigated the functional relevance of TBX20 in endocardial lineages for cardiac development. Analysis of Tie2-Cre \(^{Tbx20fl/null}\) mutants revealed previously unrecognized requirements for TBX20 in endocardial lineages for multiple aspects of cardiac septation. Although we did not observe defects in vascular patterning, we cannot formally rule out the possibility that vascular defects may have contributed to the observed phenotypes or embryonic lethality.

TBX20 regulates the endocardial proliferation and migratory behavior of endocardium-derived mesenchymal cells. We observed perturbations in several aspects of cushion development in mutant embryos. Cushion malformations and decreased proliferation of endocardium overlaying the cushions were consistent with the results obtained with Nfatc1-Cre \(^{Tbx20fl/fl}\) mutants (11), in which decreased endocardial proliferation was attributed to decreased \(\beta\)-catenin signaling via downregulation of lymphoid enhancer–binding factor 1 (Lef1), a phenomenon we also observed in Tie2-Cre \(^{Tbx20fl/null}\) mutants.

We also found that TBX20 was required for normal dispersion of endocardium-derived cushion cells. Ex vivo endocardial explant assays demonstrated that TBX20 is not required for EMT, but that mesenchymal cells require TBX20 for subsequent matrix invasion. In vivo, mutant endocardium-derived mesenchymal cells were abnormally clustered and failed to disperse and populate the forming valves in the OFT.

TBX20 in endocardium regulates cardiomyocyte migration. We found that loss of TBX20 in endocardial lineages resulted in decreased myocardialization of the OFT, indicating a disturbed crosstalk between endocardial lineages and cardiomyocytes. Myocardialization involves polarized cell movements of OFT cardiomyocytes and is required for normal OFT alignment and septation (18, 19). Decreased myocardialization in Tie2-Cre \(^{Tbx20^{fl/fl}}\) mutants probably contributes to the observed DORV phenotype.

We describe another endocardial requirement for TBX20 in DMP formation. Tie2-Cre \(^{Tbx20^{fl/fl}}\) mutants displayed a striking absence of a DMP, which is a second heart field–derived struc-
had defects in valve development, including immature atrioventricular valves and bicuspid aortic valves (24). Myh10-null mice and mice that carry a point mutation in Myh10 have defects strikingly similar to those seen in Tie2-Cre Tbx20fl/null mutants, including DORV, decreased OFT myocardialization, and defects in the fusion of atrioventricular cushions (25, 26). While the tissue-specific roles of these factors remain to be addressed, we postulate that their downregulation in Tie2-Cre Tbx20fl/null mutants probably contributes to the observed phenotypes.

Cofactors of TBX20 in endocardial lineages. Our analysis of genomic regions that were available for TF interaction through ATAC-seq in endocardial lineages and bound by TBX20 in embryonic heart revealed overrepresentation of specific TF motifs. Several of these TFs, including NKX2-5, GATA4, MEF2, and TEAD, have been previously associated with TBX20 in adult cardiomyocytes and cardiac fibroblasts (31, 34). Interestingly, motifs highly enriched near TBX20-binding sites within endocardial lineages included those recognized by the AP1 family, which contains JUN, FOS, ATF, and MAF proteins. AP1 proteins regulate a variety of processes including cell proliferation and survival, differentiation, growth, migration, and transformation. Intriguingly, conditional deletion of the API family member Jun using Tie2-Cre leads to DORV, VSDs, and valve defects (54), suggesting that perhaps JUN cooperates with TBX20 in endocardial lineages to effect normal development of endocardial cushions.

Tbx20 directly regulates the expression of ECM and migration genes. Consistent with the kinds of abnormal cellular behaviors we observed in Tie2-Cre Tbx20fl/fl mutants, microarray analysis of purified endocardial lineages showed downregulation of ECM and axon guidance genes. The intersection of this data set with TBX20 ChIP-seq data from whole embryonic heart revealed that, in mutants, several direct targets of TBX20 that have essential roles in cardiac septation were downregulated, including Robo1, Myh10, and Vcan. Robo1 mutants had VSDs similar to those seen in Tie2-Cre Tbx20fl/fl mutants, and Robo1 Robo2 compound mutants had defects in valve development, including immature atrioventricular valves and bicuspid aortic valves (24). Myh10-null mice and mice that carry a point mutation in Myh10 have defects strikingly similar to those seen in Tie2-Cre Tbx20fl/null mutants, including DORV, decreased OFT myocardialization, and defects in the fusion of atrioventricular cushions (25, 26). While the tissue-specific roles of these factors remain to be addressed, we postulate that their downregulation in Tie2-Cre Tbx20fl/null mutants probably contributes to the observed phenotypes.
septation. Both ETS and FOX factor motifs were also overrepresented in our ATAC-seq peaks, in keeping with the important role of ETS-FOX factors in endothelium-specific gene expression (55). A recent analysis of NKX2-5 mutants associated with CHD found that ETS factors are embedded in the cardiac gene–regulatory network (56). Moreover, the bioinformatics analysis of multiple DNA adenine methyltransferase identification (DAM-ID) and ChIP-seq experiments predicts a functional interaction between ETS factors and T-box factors in the regulation of cardiac target gene expression (56). Our finding that ETS motifs were highly enriched near endocardial TBX20-binding sites provides additional evidence for such interactions and further underlines the importance of TF interactions in cardiac development.

Overrepresentation of GATA TF motifs is of particular interest, as cardiac septation is especially sensitive to the dosage of GATA4, GATA5, and GATA6. Ablation of Gata4 from endocardium is embryonically lethal before E12.5 due to EMT defects in cushions (57). Additionally, a Gata4 hypomorphic allele is associated with DORV and AVSDs (58), and, interestingly, compound heterozygotes for Gata4 and Gata5 or for Gata5 and Gata6 also show DORV and VSDs (59). The endocardium-specific requirements for GATA factors remain to be addressed, but the foregoing findings suggest that TBX20 may interact with GATA factors within endocardial lineages to effect septation.

Identification of long-range enhancer targets with limited numbers of cells. Mapping enhancers to their true target genes remains challenging, especially when it concerns developmental processes that occur in small numbers of cells. In this study, we show that the intersection of whole-heart TBX20 ChIP-seq and endocardial lineage–specific assessment of open chromatin is a powerful tool for identifying putative endocardial lineage enhancers. In addition, we found that the intersection of these data sets with existing HiC data from a similar cell type (51) proved instrumental in mapping long-range–acting, cell-type–specific enhancers to their putative target gene. With the observation that chromatin architecture is normally expressed in fibroblasts may prove useful in understanding Vcan regulation in the context of tumorigenesis and could provide a therapeutic target.

Methods

Mouse strains. Mice were maintained on a Black Swiss (NIHBL(S); Tac- tonic Biosciences) outbred background. Tbx20-conditional and -null alleles and the Tbx20-GFP and Nfatc1-Cre mouse lines were generated in the authors’ laboratories and have been previously described (8, 16, 31). Tie2-Cre (69), Rosa26 flox-stop-flox tdTomato (Rosa tdTom) (70), and Rosa26 membrane-targeted (m) tdTom flox-stop-flox mGFP (Rosa mT/mGFP) (71) mice were obtained from The Jackson Laboratory.

Histology. Embryos were isolated in PBS and fixed in 4% paraformaldehyde in PBS at 4°C for 6 to 12 hours. Tissue was dehydrated using ethanol (50%, 70%, 80%, 90%, 96%, 3 × 100%; 5 minutes each), cleared in Histo-Clear (National Diagnostics) for 30 minutes, incubated 3 times for 30 minutes in Paraplast X-tra (Leica Biosystems), and embedded for sectioning. Alternatively, tissue was dehydrated in a sucrose gradient (5%, 10%, 15%, 20%, 6–12 hours each), embedded in 20% sucrose:OCT, and frozen for cryosectioning. Sections (10- to 12-μm thick) were used for H&E staining or immunostaining, as described previously (31).

Cushion explant assay. E10.5 OFT cushions were dissected and cultured on collagen matrices as described previously (72). After 72 hours, explants were fixed for 30 minutes in 4% PFA, stained with phalloidin-647 and DAPI, and imaged using a Leica TCS SP5 confocal microscope. Quantifications were performed on Tie2-Cre lineage–traced cells.

Cell quantification experiments. Cell-counting experiments were performed using Velocity Software (PerkinElmer) or manually using ImageJ (NIH). In vivo quantification of cell numbers and EdU labeling were performed by counting all lineage-traced cushion cells in every other section of E11.5 hearts. Cell-counting experiments in ex vivo culture explants were performed by manually counting all lineage-traced mesenchymal cells that migrated away from the explant over the surface of the collagen gel, using a Leica SP5 confocal microscope. Quantification of invasion was performed using differential interference microscopy, counting all Tie2-Cre lineage–traced cells at marked distances beneath the surface of the collagen gel in

A recent analysis of NKX2-5 mutants associated with CHD found that ETS factors are embedded in the cardiac gene–regulatory network (56). Moreover, the bioinformatics analysis of multiple DNA adenine methyltransferase identification (DAM-ID) and ChIP-seq experiments predicts a functional interaction between ETS factors and T-box factors in the regulation of cardiac target gene expression (56). Our finding that ETS motifs were highly enriched near endocardial TBX20-binding sites provides additional evidence for such interactions and further underlines the importance of TF interactions in cardiac development.

Overrepresentation of GATA TF motifs is of particular interest, as cardiac septation is especially sensitive to the dosage of GATA4, GATA5, and GATA6. Ablation of Gata4 from endocardium is embryonically lethal before E12.5 due to EMT defects in cushions (57). Additionally, a Gata4 hypomorphic allele is associated with DORV and AVSDs (58), and, interestingly, compound heterozygotes for Gata4 and Gata5 or for Gata5 and Gata6 also show DORV and VSDs (59). The endocardium-specific requirements for GATA factors remain to be addressed, but the foregoing findings suggest that TBX20 may interact with GATA factors within endocardial lineages to effect septation.

Identification of long-range enhancer targets with limited numbers of cells. Mapping enhancers to their true target genes remains challenging, especially when it concerns developmental processes that occur in small numbers of cells. In this study, we show that the intersection of whole-heart TBX20 ChIP-seq and endocardial lineage–specific assessment of open chromatin is a powerful tool for identifying putative endocardial lineage enhancers. In addition, we found that the intersection of these data sets with existing HiC data from a similar cell type (51) proved instrumental in mapping long-range–acting, cell-type–specific enhancers to their putative target gene. With the observation that chromatin architecture is remarkably well conserved between cell types and between species (60, 61), we predict that our approach can be broadly applied to other systems, working with limited numbers of cells.

TBX20 regulates Vcan expression in endocardial and mesenchymal cells via a distant enhancer. The intersection of our data sets with HiC data suggested that Vcan, downregulated in Tie2-Cre Tbx20−/− mutant endocardial lineages, might have long-range enhancers that were directly targeted by TBX20. This was of particular interest, as mice homozygously mutant for hypomorphic alleles of Vcan have phenotypes, including hypomorphic atrioventricular cushions, reduced DMP, and lack of myocardialization of Vcan.

Our analysis of distant enhancers in the Vcan locus identified lineage-specific enhancers that drive expression in cells that normally express Vcan, strongly suggesting that we have successfully identified cardiac enhancers for Vcan. These enhancers are likely to be functionally conserved between species, as chromatin interactions between the Vcan enhancers we identified and the Vcan TSSs were found in human and mouse HiC experiments (51, 60). Furthermore, the enhancers have epigenetic marks of active enhancers in human hearts. Taken together, our results provide evidence that the regulation of Vcan by TBX20 in endocardial lineages is conserved between humans and mice and that VCAN misregulation in patients with TBX20 mutations may contribute to CHDs such as DORV or AVSD (3, 6).

High expression levels of VCAN have been found in most malignancies and are associated with cancer relapse and poor patient outcomes in numerous cancer types, including breast and prostate cancers (63). VCAN is predominantly secreted by activated peritumoral fibroblasts and vasculature and has also been reported to be secreted by some cancer cells (63, 64). Inhibition of VCAN expression or function in glioma cells, ovarian cancer cells, or lung cancer cells has been shown to inhibit cell invasion (64–66). Targeting of VCAN thus provides a potential strategy for suppressing tumor invasion (65, 67, 68). Our identification of a transcriptional enhancer that is active in fibroblasts may prove useful in understanding Vcan regulation in the context of tumorigenesis and could provide a therapeutic target.

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1 field of view (×5 objective), which typically represents one-quarter of the entire explant. Migration distance was defined as the distance between the edge of the explant and the cell body. The number of biological replicates used for these experiments was 3 or more and is reported in the figure legends.

Abs. Abs used for immunostaining included GFP (Ab13790; 1:1,000; Abcam); ACTN2 (A7811; 1:200; Sigma-Aldrich); TNNT2 (Ab106076; 1:2,000; Abcam); cleaved caspase-3 (9664; 1:200; Cell Signaling Technology); VCAN (Ab1033; 1:200; EMD Millipore); and CD31 (73117; 1:50; BD Biosciences). EdU (100 μl, 3 g/l) was injected i.p. into female mice 2 hours prior to isolation of the embryos. EdU staining (C10337; Molecular Probes) was performed according to the manufacturer’s instructions. The following FACS Abs were obtained from eBioscience: CD31-APC (17-0311-82; 1:50); TIE2-PE (12-5987-81; 1:50); CD41 PE-Cy7 (25-0411-82; 1:100); CD45 PE-Cy7 (25-0451-82; 1:100); and Ter119 PE-Cy7 (25-5921-82; 1:100).

FACS sorting. Hearts were harvested in ice-cold PBS with 0.5 mM EDTA and dissociated as described previously (73), using collagenase (10 mg/ml; Worthington) and dispase (10 mg/ml, Invitrogen). Cells were stained for 30 minutes, and DAPI was used to label dead cells. Tie2-Cre lineage cells, negative for the blood markers CD41, CD45, and Ter119, were sorted on an Influx Cell Sorter (BD Biosciences) and collected in RNA lysis buffer (Agilent) or TRIzol reagent (Ambion) for RNA extraction, or in PBS and 5% serum for ATAC-seq.

RNA extraction, microarray analysis, and qRT-PCR. RNA was isolated from 10,000 FACS-sorted cells using the Agilent RNA Nanoprep Kit. Amplification, hybridization to the Affymetrix Mouse Genome 430 2.0 Array GeneChip, and analysis were performed as previously described (75). Amplification, hybridization to the Affymetrix Mouse Genome 430 2.0 Array GeneChip, and analysis were performed as previously described (75). The following FACS Abs were obtained from eBioscience: CD31-APC (17-0311-82; 1:50); TIE2-PE (12-5987-81; 1:50); CD41 PE-Cy7 (25-0411-82; 1:100); CD45 PE-Cy7 (25-0451-82; 1:100); and Ter119 PE-Cy7 (25-5921-82; 1:100).

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RNA extraction, microarray analysis, and qRT-PCR. RNA was isolated from 10,000 FACS-sorted cells using the Agilent RNA Nanoprep Kit. Amplification, hybridization to the Affymetrix Mouse Genome 430 2.0 Array GeneChip, and analysis were performed as previously described (73). Genes “present” in controls and downregulated or “present” in mutants and upregulated by 1.5-fold or more were considered differentially expressed. Data are available in the NCBI’s Gene Expression Omnibus (GEO) database (GEO GSE73858). For qRT-PCR, RNA was extracted from sorted cells using TRIzol, and cDNA was generated using SuperScript VILO (Invitrogen). qRT-PCR was performed using FastStart SYBR Green Master Mix (Roche) on a Bio-Rad CFX96 Real-Time PCR System and the primers listed in Supplemental Table 5.

TBX20 ChIP-seq. E11.5 TBX20-GFP mouse hearts were dissected and cross-linked in 1.8% formaldehyde for 10 minutes at room temperature, quenched in 2.5 M glycine for 5 minutes, washed, and resuspended in lysis buffer (1% SDS, 10 mM EDTA, 50 mM Tris, pH 8.1). Cross-linked chromatin was sonicated using a Misonix 4000 (amplitude of 90; 30 seconds on/30 seconds off) to an average size of 8.1). Cross-linked chromatin extract was incubated overnight at 4°C with goat anti-GFP and immunoprecipitated with protein A sepharose beads. DNA (10 ng) was used to generate a standard Illumina sequencing library. Sequencing reads were aligned to mm9 using cutadapt (75). Paired-end alignment was performed with bowtie2 2.2.3, with a maximum distance of 2 kb between mates. Peaks were called using HOMER (32) against whole-heart input with a 1 × 10⁻⁶ FDR cutoff. Data are available in the EMBL-EBI database under accession number E-MTAB-3972.

Vcan enhancer in vivo reporter assays. Vcan enh1 (Chrl3: 9097306-90973914) and Vcan enh2 (Chrl3: 91025841-91026597) were cloned into Hsp68-LacZ (76), and mouse in vivo transgenic reporter assays were performed by pronuclear injection as reported previously (77). TBX20 motifs in Vcan enh1 were modified by Gene Tailor site-directed mutagenesis (Invitrogen) using the primers listed in Supplemental Table 5. Embryos were harvested at E12.5 and subjected to X-gal staining as reported previously (77). Embryos were genotyped using LacZ primers (forward, 5′-CADCCTGAAATGGCGAATGGCGCTT; reverse, 5′-CCGGTTGCCACCCAGATGAAAGCC), and embryos that were (nearly) completely stained blue (between 1 and 3 for each line) were excluded from further analysis.

Statistics. Values are presented as the mean ± SEM. For ex vivo cell migration assays, a nonpaired, 2-tailed t test was used to analyze the migration distance between controls and mutants. Matrix invasion was analyzed using 2-way ANOVA, with Bonferroni’s post tests. An unpaired, 2-tailed t test was used to analyze qRT-PCR data. A P value of less than 0.05 was considered significant.

Study approval. Animal care and experimental procedures were performed according to protocols approved by the IACUCs of the UCSD protocol 504150 and the University of Chicago (protocol 71656).

Author contributions

CJB, IA, NS, RJD, QJC, JC, MAN, and SME designed and/or performed experiments and analyzed the data. CJB, IA, NS, MAN, and SME wrote the manuscript. BZ provided reagents.

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