Mutations in tropomyosin 4 underlie a rare form of human macrothrombocytopenia

Irina Pleines, …, Marloes R. Tijssen, Benjamin T. Kile


Platelets are anuclear cells that are essential for blood clotting. They are produced by large polyploid precursor cells called megakaryocytes. Previous genome-wide association studies in nearly 70,000 individuals indicated that single nucleotide variants (SNVs) in the gene encoding the actin cytoskeletal regulator tropomyosin 4 (TPM4) exert an effect on the count and volume of platelets. Platelet number and volume are independent risk factors for heart attack and stroke. Here, we have identified 2 unrelated families in the BRIDGE Bleeding and Platelet Disorders (BPD) collection who carry a TPM4 variant that causes truncation of the TPM4 protein and segregates with macrothrombocytopenia, a disorder characterized by low platelet count. N-Ethyl-N-nitrosourea–induced (ENU-induced) missense mutations in Tpm4 or targeted inactivation of the Tpm4 locus led to gene dosage–dependent macrothrombocytopenia in mice. All other blood cell counts in Tpm4-deficient mice were normal. Insufficient TPM4 expression in human and mouse megakaryocytes resulted in a defect in the terminal stages of platelet production and had a mild effect on platelet function. Together, our findings demonstrate a nonredundant role for TPM4 in platelet biogenesis in humans and mice and reveal that truncating variants in TPM4 cause a previously undescribed dominant Mendelian platelet disorder.
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Irina Pleines,1,2 Joanne Woods,3,4 Stephane Chappaz,1,2 Verity Kew,3,4 Nicola Foad,3,4 José Ballester-Beltrán,3,4 Katja Aurbach,5 Chiara Lincetto,3,4 Rachael M. Lane,1 Galina Schevzov,4 Warren S. Alexander,1,2 Douglas J. Hilton,1,2 William J. Astle,3,4 Kate Downes,3,4 Paquita Nurden,7 Sarah K. Westbury,8 Andrew D. Mumford,8 Samya G. Obaji,10 Peter W. Collins,10 NIHR BioResource–Rare Diseases, Cambridge University Hospitals, Cambridge Biomedical Campus, Cambridge, United Kingdom. 12 Transgenic Animal Unit, Mark Wainwright Analytical Centre, University of Cambridge, Cambridge, United Kingdom. 13 Arthur Bloom Haemophilia Centre, School of Medicine, Cardiff University, Heath Park, Cardiff, United Kingdom. 11 Walter and Eliza Hall Institute of Medical Research, Parkville, Australia. 12 Department of Haematology, University of Cambridge, Cambridge Biomedical Campus, Cambridge, United Kingdom. 14 Institute of Experimental Biomedicine, University Hospital and Rudolf Virchow Center, University of Wuerzburg, Wuerzburg, Germany. 5 School of Medical Sciences, University of New South Wales, Sydney, Australia. 6 Institut Hospitalo-Universitaire LIRYC, Plateforme Technologique d’Innovation Biomédicale, Hôpital Xavier Amoison, Pessac, France. 7 School of Clinical Sciences, University of Bristol, Bristol, United Kingdom. 8 School of Cellular and Molecular Medicine, University of Bristol, Bristol, United Kingdom. 9 Arthur Bloom Haemophilia Centre, School of Medicine, Cardiff University, Heath Park, Cardiff, United Kingdom. 10 NIHR BioResource–Rare Diseases, Cambridge University Hospitals, Cambridge Biomedical Campus, Cambridge, United Kingdom. 11 Transgenic Animal Unit, Mark Wainwright Analytical Centre, University of New South Wales, Sydney, Australia. 12 Human Genetics, Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, United Kingdom. 13 Medical Research Council Biostatistics Unit, Cambridge Institute of Public Health, Cambridge, United Kingdom.

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Introduction

Platelets are small, anuclear blood cells that are essential for blood clotting and the maintenance of vascular integrity (1, 2). They are produced by megakaryocytes, large polyploid precursor cells that develop primarily in the bone marrow. Megakaryocytes undergo a unique process of growth and maturation involving replication of DNA without cell division (endomitosis), a massive expansion of cytoplasmic volume, and the development of an extensive internal membranous network known as the demarcation membrane system. Upon maturity, megakaryocytes form filamentous protrusions termed “proplatelets” that are extruded into the bone marrow sinusoids. Evidence suggests platelet biogenesis is the result of individual platelets budding off from proplatelets, as well as proplatelets themselves being shed and subsequently maturing into platelets within the circulation (3). The molecular regulation of this unique process of blood cell formation is only partially understood.

The number, cellular volume, and function of platelets are tightly regulated within individuals. However, all 3 parameters vary widely at the population level, and represent independent risk factors for heart attack and stroke (4). Population variation is to a large extent explained by common single nucleotide variants (SNVs), which exert small effects. Major deviations are triggered by rare variants that have effects of sufficient magnitude to cause Mendelian platelet disorders. A metaanalysis of genome-wide association studies identified 68 independent common SNVs associated with platelet count and volume (5). Several of these SNVs were linked to genes encoding proteins with well-established roles in platelet formation (e.g., THPO [thrombopoietin], GPIBA [glycoprotein Ibα], TUBBI [tubulin-β1]) and survival (the apoptosis regulator BAK [encoding BAK]). However, the vast majority of identified SNVs were located near genes hitherto uncharacterized in the megakaryocyte lineage.

Authorship note: M.R. Tijssen and B.T. Kile contributed equally to this work.
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Tropomyosins, which form copolymers with actin, are a prime example, with SNVs rs3809566 and rs8109288 marking the genes encoding tropomyosin 1 (TPM1) and tropomyosin 4 (TPM4), respectively. It has been reported that expression of transcripts from the TPM4 locus strongly increases in the later stages of megakaryopoiesis (5, 6). In addition, chromatin immunoprecipitation in primary megakaryocytes for 5 transcription factors with essential roles in platelet biogenesis revealed that TPM4 contains a nucleosome-depleted regulatory element occupied by all 5 factors (7), placing the gene in a unique category of fewer than 200 genes. Morpholino knockdown of its homolog tpma in zebrafish abolished the formation of thrombocytes, suggesting a functional role for TPM4 in mammalian thrombopoiesis (5, 8).

Tropomyosins are ubiquitously expressed and highly conserved. Mouse and human homologs exhibit greater than 90% identity at the amino acid level. In mammals, tropomyosins are encoded by 4 different genes: TPM1, TPM2, TPM3, and TPM4. Alternative promoter usage and alternative splicing result in approximately 40 different TPM protein isoforms in humans (reviewed in ref. 9; for nomenclature used in this study, see ref. 10). Intriguingly, despite their similarity, individual tropomyosin genes have largely nonredundant functions, as illustrated by the fact that the constitutive deletion of either Tmp1, Tmp2, or Tmp3 in mice results in embryonic lethality (11–13). Tmp4 knockout mice have not previously been described.

The best-characterized tropomyosins are the muscle-specific isoforms, which mediate the interaction between myosin and actin. In humans, mutations affecting the expression of muscle tropomyosins cause familial hypertrophic cardiomyopathy (TPM1) and nemaline myopathies (TPM2 and TPM3). These disease phenotypes are recapitulated in mice carrying similar mutations (14). The “nonmuscle” (or “cytoskeletal”) tropomyosin isoforms are often referred to as “gatekeepers,” as they regulate the accession of other actin-binding proteins and myosin motors to the actin filament (9). The cytoskeletal tropomyosins are implicated in the regulation of cell morphology, adhesion, migration, granule trafficking, cell division, and apoptosis. Except for an involvement in modulating erythrocyte membrane stability (15), virtually nothing is known about the roles of tropomyosins in hematopoietic cells.

Here, we demonstrate that, in both humans and mice, TPM4 is an essential mediator of platelet biogenesis. Genome sequencing of 865 index cases with bleeding and platelet disorders of unknown molecular etiology revealed 2 pedigrees with a premature stop codon in TPM4. This variant was absent from nearly 75,000 control DNA samples and segregated with large platelets and concomitant low platelet count (a phenotype termed macrothrombocytopenia) in the 2 pedigrees. Mutation of murine Tmp4 resulted in dose-dependent macrothrombocytopenia caused by a defect in the terminal stages of platelet production. This was recapitulated by shRNA knockdown in human megakaryocytes. In addition to a reduction in platelet number and an increase in mean platelet volume, a mild defect in platelet function was observed in both humans and mice. Our findings demonstrate that TPM4 encodes a tropomyosin with an important nonredundant role in platelet biogenesis, and define a novel, dominant, Mendelian platelet disorder.

**Results**

**Tropomyosin expression in human and mouse megakaryocytes and platelets.** Previous RNA sequencing studies have indicated that human megakaryocytes express TPM1, TPM3, and TPM4, and low levels of TPM2 (ref. 6 and Supplemental Figure 1A; supplemental material available online with this article; https://doi.org/10.1172/JCI86154DS1). We performed reverse transcriptase–PCR (RT-PCR), which confirmed these findings, and demonstrated that TPM4 is the most highly expressed of the tropomyosins, followed by TPM1 (Figure 1A). Western blotting of whole cell lysates demonstrated the presence of protein products of all 4 tropomyosin genes in WT mouse and healthy human control platelets (Figure 1B). The short TPM4 30-kDa isoform (TPM4.2) was the major isoform in both species. Human platelets additionally appear to harbor small amounts of the long 38-kDa TPM4 isoform (TPM4.1). Hereafter, the term “TPM4” is used to indicate both protein isoforms TPM4.1 and TPM4.2 (in humans) or solely TPM4.2 (in mice).

**A truncating mutation in TPM4 causes macrothrombocytopenia in humans.** We next performed a look-up in the database for the Biomedical Research Centres/Units Inherited Diseases Genetic Evaluation (BRIDGE) Bleeding and Platelet Disorders (BPD) collection. To date, over 1,000 participants have been enrolled and genome-sequenced as part of the BRIDGE-BPD study for genome sequencing along with approximately 5,000 participants with other rare diseases enrolled to the NIHR BioResource–Rare Diseases Biomedical Research Centres/Units Inherited Diseases Genetic Evaluation (ExAC) database (17) as well as the UK10K database (18). The 2 cases do not share any other rare variants, and their estimated genome-wide kinship, based on common unlinked SNVs (19), was estimated to be zero, suggesting that the variant did not arise in a recent common ancestor.

Predicted loss-of-function variants in TPM4.2 are rare in the general population, with only 2 heterozygotes observed among the
above 13 fl or if there was presence of macrothrombocytes as observed by electron microscopy. The propositus of pedigree 1 carrying the premature stop codon was a woman of mixed European descent who was enrolled to the BRIDGE-BPD study at 56 years old with a history of mild bleeding (see Supplemental Methods for full clinical history) consistent with observed platelet counts in the range of 80 × 10^9 to 110 × 10^9 per liter. Other than breast carcinoma, there was no other relevant past medical history. Analysis of 2 more than 60,000 individuals in ExAC (1 with a splice site variant and 1 with a variant encoding a premature stop at residue 54). The variant concerning the BPD patients encodes a premature stop codon in exon 3 of TPM4, resulting in a truncation at residue 69 of the major protein isoform TPM4.2, and residue 105 of the minor isoform TPM4.1 (Figure 2A).

Macrothrombocytopenia was deemed present if the platelet count was below 150 × 10^9 per liter or the mean platelet volume (MPV) was above 13 fl or if there was presence of macrothrombocytes as observed by electron microscopy. The propositus of pedigree 1 carrying the premature stop codon was a woman of mixed European descent who was enrolled to the BRIDGE-BPD study at 56 years old with a history of mild bleeding (see Supplemental Methods for full clinical history) consistent with observed platelet counts in the range of 80 × 10^9 to 110 × 10^9 per liter. Other than breast carcinoma, there was no other relevant past medical history. Analysis of 2
siblings and the propositus’ daughter demonstrated segregation of the observed variant with macrothrombocytopenia (Figure 2B and Supplemental Figure 1B). The propositus (1-II-5) and her daughter (1-III-7) exhibited platelet counts lower than those of the unaffected siblings (103 × 10⁹ and 140 × 10⁹ per liter compared with 226 × 10⁹ and 236 × 10⁹ per liter, respectively) with mean platelet volumes well above 13 fl. All other blood counts were within normal range (Table 1), suggesting a specific role for TPM4 in the megakaryocyte lineage.

The propositus of pedigree 2 was a woman of mixed European descent enrolled in the BRIDGE-BPD study at 58 years of age. She had a platelet count of 230 × 10⁹ per liter with a platelet volume near the upper limit of normal (12.9 fl). MacrORTHOMBocytes were observed by electron microscopy. She had suffered with menorrhagia from menarche that did not improve upon hormonal therapy and therefore required recurrent dilatation and curettages, and finally a hysterectomy at the age of 37 (see Supplemental Methods for full clinical history). Platelet function tests including von Willebrand parameters and coagulation factors (IX, XI, and XIII) revealed no abnormality. Other than severe asthma and eczema, there was no past medical history of note. Analysis of 3 of the propositus’ children demonstrated segregation of the observed variant with a lower platelet count and larger platelets (Figure 2B and Supplemental Figure 1C). Pedigree member 2-III-5, who carries the variant, had a platelet count and mean platelet volume just within the observed variant with macrothrombocytopenia (Figure 2B and Supplemental Figure 1C). Pedigree member 2-III-5, who carries the variant, had a platelet count and mean platelet volume just within the normal range (160 × 10⁹ and 12.3 fl, respectively). He experienced no abnormal bleeding in response to a number of hemostatic challenges, including a laparotomy, orchiopexy, and 4 dental extractions. His sibling (2-III-7), who also carries the mutation, exhibited a low platelet count (146 × 10⁹ per liter) and large mean platelet volume (14.6 fl), and a bleeding tendency similar to the propositus’ (see Supplemental Methods for full clinical history). 2-III-6 does not carry the variant and had the highest platelet count (278 × 10⁹ per liter) and lowest mean platelet volume (10.8 fl) of all family members. Thus, the variant cosegregates with isolated macrothrombocytopenia.

TPM4.2 mRNA levels were reduced to 59% and 78% of the control in the affected cases 1-II-5 and 1-III-7, respectively (Figure 2C), and a corresponding approximately 50% reduction in the expression of TPM4.2 protein was observed with an antibody raised against the TPM4 C-terminus (δ/9d) (Figure 2D). Pedigree 2 exhibited similar defects, with mRNA levels reduced to 46%–63%, and we were able to detect a corresponding reduction in TPM4.2 protein in 2 of 3 affected pedigree members (Supplemental Figure 1D). The sex-stratified quantiles of mean platelet volume measurements with respect to the INTERVAL study (measurements obtained using a Sysmex hematology analyzer from 48,345 blood donors) were all above 91%, with 3 of the 5 measurements at percentiles exceeding 99.9% (20). This indicates that the platelets of the carriers are consistently large, with some extremely large. The sex-stratified quantiles of the platelet count (PLT) measurements with respect to INTERVAL were below 4% in all cases except the index case of pedigree 2, with 3 of the 5 measurements at percentiles below 1%. In contrast, all the WT homozygotes had PLT quantiles between 23% and 82% (Figure 2E). The probability of observing cosegregation under the null hypothesis that the mutation is not causal, conditional on having already observed the mutant in the propositus, is 0.5¹ for each pedigree, and P = 0.0156 in total. Collectively, these data provide convincing evidence that loss-of-function variants in TPM4 can cause macrothrombocytopenia, and suggest it plays a role in the development of the megakaryocyte lineage.

A loss-of-function allele of murine Tpm4 causes macrothrombocytopenia. In parallel studies, we conducted an N-ethyl-N-nitrosourea (ENU) mutagenesis screen in mice for mutations causing dominant thrombocytopenia (21, 22). A mouse exhibiting a decrease in platelet number and increase in mean platelet volume was identified and named Plt53. Fifty percent of the progeny from this animal also exhibited macrothrombocytopenia. After backcrossing, exome sequencing of 2 affected animals followed by genotyping of candidate mutations in siblings and offspring revealed that the Plt53 phenotype correlated with inheritance of a point mutation in the gene encoding TPM4. The mutation, at nucleotide g.72,147,268, was a substitution of A for G in the first nucleotide dinucleotide of the donor splice site in exon 7 of the Tpm4 gene (hereafter referred to as Tpm4<sup>Plt53/+</sup>). RT-PCR analysis of cDNA extracted from megakaryocytes revealed that the Plt53 mutation led to the inclusion of part of intron 7 in the Tpm4 mRNA. The predicted result is a protein bearing 8 intron-encoded amino acids followed by premature truncation due to the presence of an in-frame stop codon at nucleotide g.72,147,291 (Figure 3A).

Western blotting with an antibody raised against the TPM4 C-terminus (δ/9d) revealed the complete absence of TPM4 in Tpm4<sup>Plt53/Plt53</sup> platelets, and reduced expression in Tpm4<sup>Plt53/+</sup> platelets (Figure 3B, top). The lower band observed with this antibody was not detected with 2 other TPM4 antibodies and is therefore most likely unspecific (Supplemental Figure 2A). Using an N-ter-

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**Table 1. Blood counts of cases of families 1 and 2 carrying the TPM4 variant, including the depiction of normal count variation**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>1-II-5</th>
<th>1-III-7</th>
<th>2-II-3</th>
<th>2-III-5</th>
<th>2-III-6</th>
<th>2-III-7</th>
<th>Normal range</th>
</tr>
</thead>
<tbody>
<tr>
<td>rbc (×10⁹/liter)</td>
<td>4.72</td>
<td>4.52</td>
<td>4.84</td>
<td>5.29</td>
<td>5.50</td>
<td>4.68</td>
<td>3.80–4.80</td>
</tr>
<tr>
<td>HCT (%)</td>
<td>42</td>
<td>41</td>
<td>45</td>
<td>49</td>
<td>49</td>
<td>43</td>
<td>37–45</td>
</tr>
<tr>
<td>Lymphocytes (×10⁹/liter)</td>
<td>1.31</td>
<td>1.82</td>
<td>1.4</td>
<td>1.4</td>
<td>1.8</td>
<td>1.5</td>
<td>1.00–4.00</td>
</tr>
<tr>
<td>Neutrophils (×10⁹/liter)</td>
<td>3.68</td>
<td>4.67</td>
<td>3.9</td>
<td>2.9</td>
<td>3.0</td>
<td>7.9</td>
<td>1.50–7.50</td>
</tr>
<tr>
<td>Basophils (×10⁹/liter)</td>
<td>0.03</td>
<td>0.02</td>
<td>0.1</td>
<td>0.0</td>
<td>0.1</td>
<td>0.1</td>
<td>&lt;0.11</td>
</tr>
<tr>
<td>Monocytes (×10⁹/liter)</td>
<td>0.24</td>
<td>0.7</td>
<td>0.4</td>
<td>0.6</td>
<td>0.4</td>
<td>0.7</td>
<td>0.20–1.00</td>
</tr>
<tr>
<td>Eosinophils (×10⁹/liter)</td>
<td>0.07</td>
<td>0.05</td>
<td>0.3</td>
<td>0.1</td>
<td>0.1</td>
<td>0.1</td>
<td>&lt;0.51</td>
</tr>
<tr>
<td>Neutrophils (×10⁹/liter)</td>
<td>0.24</td>
<td>0.7</td>
<td>0.4</td>
<td>0.6</td>
<td>0.4</td>
<td>0.7</td>
<td>0.20–1.00</td>
</tr>
<tr>
<td>Eosinophils (×10⁹/liter)</td>
<td>0.07</td>
<td>0.05</td>
<td>0.3</td>
<td>0.1</td>
<td>0.1</td>
<td>0.1</td>
<td>&lt;0.51</td>
</tr>
</tbody>
</table>

Measurements obtained using a Sysmex hematology analyzer. HCT, hematocrit.
minal specific antibody (δ/1b), we observed a faint band of lower molecular weight in platelets from heterozygous and homozygous mutant mice, indicating the presence of some residual truncated TPM4 protein (Figure 3B, middle). Given that the overlap of N- and C-termini of TPM dimers is essential for filament formation (23), these data indicated that the Plt53 allele of Tpm4 represents a complete loss of the protein’s canonical function.

Heterozygous (Tpm4<sup>Plt53/+</sup>) and homozygous (Tpm4<sup>Plt53/Plt53</sup>) animals appeared healthy, and histopathological examination revealed no obvious organ abnormalities or pathology. Relative to WT littermates, Tpm4<sup>Plt53/+</sup> mice exhibited mild macrothrombocytopenia (Figure 3C), and Tpm4<sup>Plt53/Plt53</sup> animals a more severe form, with approximately 50% reduction in platelet counts. Remarkably, other peripheral blood cell counts in Tpm4<sup>Plt53/+</sup> and Tpm4<sup>Plt53/Plt53</sup> mice were unchanged, with the exception of a modest, but statistically significant (P = 0.0008), increase in lymphocyte counts in the Tpm4<sup>Plt53/Plt53</sup> cohort (Table 2). Flow cytometric analysis of bone marrow revealed normal numbers of immunophenotypic hematopoietic progenitor cells, including megakaryocyte progenitors, in Tpm4<sup>Plt53/Plt53</sup> mice (Table 3). Transplantation of unfractionated bone marrow from Tpm4<sup>Plt53/Plt53</sup> animals into lethally irradiated adult recipients resulted in macrothrombocytopenia (Supplemental Figure 2B), demonstrating that the phenotype was intrinsic to the hematopoietic system. To confirm the Plt53 mutation as the causative genetic lesion, we generated an independent knockout mouse strain for TPM4.2, the only TPM4 protein isoform expressed in mice. Strikingly, given the early lethality observed in Tpm1, Tpm2, and Tpm3 knockouts, animals lacking Tpm4 were viable and outwardly healthy. They exhibited a dose-dependent macrothrombocytopenia (Figure 3D and Supplemental Figure 3,
Lymphocytes (×10^3/ml) 8.7 ± 1.7 9.1 ± 1.0 (NS) 10.7 ± 1.5
HCT (%) 62.6 ± 3.0 63.7 ± 2.2 (NS) 62.6 ± 1.9 (NS)

A–C), similar in magnitude to that observed in Tpm4^−/− mutants on a C57BL/6 background (the background on which the targeted knockout allele was generated) (Supplemental Figure 3D). Together, these data demonstrate a specific requirement for TPM4 in the megakaryocyte lineage.

**TPM4 insufficiency results in aberrant platelet morphology.** Transmission electron microscopy (TEM) analysis of platelets from the 2 affected cases from pedigree 1 revealed considerable heterogeneity in platelet size, some of them being very large. We also observed an increased presence of large vacuoles (Figure 4, A and B). Blinded analysis of TEM pictures correctly identified all the carriers from pedigree 2. Platelets from 2-II-3 and 2-III-5 exhibited moderate defects (Supplemental Figure 4) similar to those observed for pedigree 1 (Figure 4, A and B). Platelets from 2-III-7 were obviously different from the control, with an increased small diameter and an ellipsoid rather than discoid shape.

Consistent with the human data, TEM analysis revealed considerable heterogeneity in the size and morphology of Tpm4^Plt53/Plt53 and Tpm4^Plt53/− platelets, with frequent vacuoles, indicating increased fragility of the mutant platelets (Figure 4C). Given that ultra-rare variants in FLNA (the gene encoding filamin A) have recently been reported to result in fragile platelets and macrothrombocytopenia (24), and variants in the actin–cross-linking protein actinin 1 (α-actinin, ACTN1) are one of the most frequent causes of isolated thrombocytopenia with altered platelet morphology (25–27), we examined the levels of these 2 proteins. Interestingly, we found that filamin A was partially degraded in Tpm4^Plt53/Plt53 and to a lesser extent Tpm4^Plt53/− platelets (Figure 4D, top). Levels of full-lengthactin1were reduced in Tpm4^Plt53/Plt53 and Tpm4^Plt53/− platelets, coinciding with the appearance of an 80-kDa band that has been previously reported to be the result of cleavage by calpain (Figure 4D, bottom) (28).

**Reduced TPM4 expression impacts platelet function.** We next examined the function of the platelets of the daughter of the propositor of pedigree 1 (case 1-III-7 in Figure 2B). In response to any of the agonists ADP, cross-linked collagen-related peptide (CRP-XL), and TRAP-6, no significant differences in platelet activation measured by integrin αIIbβ3 activation and α-granule secretion were revealed (Figure 5A). However, when thrombus formation under flow ex vivo was examined, both the number of thrombi and the area covered were reduced in case 1-III-7 relative to the control (P < 0.001) (Figure 5, B and C).

We then probed the impact of TPM4 dysfunction on platelet function in mice. Given that macrothrombocytopenia can be triggered by accelerated platelet clearance, we firstly examined the in vivo lifespan of Tpm4^+/+, Tpm4^Plt53/+, and Tpm4^Plt53/Plt53 platelets. No change in the kinetics of clearance or ultimate lifespan was observed, indicating that lack of functional TPM4 did not affect platelet turnover (Figure 6A). Platelet activation was assessed in vitro by flow cytometry using integrin αIIbβ3 activation (JON/A antibody) and α-granule secretion (anti–P-selectin antibody) as markers. Tpm4^Plt53/+ and Tpm4^Plt53/Plt53 platelets were responsive to a range of agonists, indicating that the processes of integrin inside-out activation and degranulation are functional in the absence of TPM4 (Figure 6B and Supplemental Figure 5A). The increased mean fluorescence intensity for integrin αIIbβ3 in the Tpm4^Plt53/Plt53 samples was most probably due to the increased size of the mutant platelets, as it correlated with an approximately 37% increase in basal CD41 (integrin αIIbβ3 subunit) fluorescence signal (Figure 6B and Supplemental Table 1). To investigate the involvement of TPM4 in activation-induced morphological changes, we incubated thrombin-activated Tpm4^+/+ and Tpm4^Plt53/Plt53 platelets on fibrinogen-coated coverslips. Under these conditions, platelets from both genotypes fully spread, but the spreading process was significantly delayed in the mutant (Figure 6C). As seen with case 1-III-7, thrombus formation on collagen under flow ex vivo was significantly reduced in blood from Tpm4^Plt53/Plt53 mice (Figure 6D and Supplemental Figure 5B). Finally, we found that tail bleeding times were mildly extended in Tpm4^Plt53/Plt53 mice, and in lethally irradiated C57BL/6 mice with transplanted Phl53/Phl53 bone marrow, relative to their respective controls (P = 0.165 and P = 0.045; Figure 6E and Supplemental Figure 5C). Together, these results demonstrate that the hemostatic function of TPM4-deficient platelets is mildly compromised.

### Table 2. Blood counts of adult Tpm4^+/+, Tpm4^Plt53/−, and Tpm4^Plt53/Plt53 mice

<table>
<thead>
<tr>
<th>Parameter</th>
<th>+/-</th>
<th>+/-/Plt53</th>
<th>Plt53/Plt53</th>
</tr>
</thead>
<tbody>
<tr>
<td>rb (×10^3/ml)</td>
<td>11.8 ± 0.4</td>
<td>12.0 ± 0.4 (NS)</td>
<td>11.6 ± 0.3 (NS)</td>
</tr>
<tr>
<td>HCT (%)</td>
<td>62.6 ± 3.0</td>
<td>63.7 ± 2.2 (NS)</td>
<td>62.6 ± 1.9 (NS)</td>
</tr>
<tr>
<td>Lymphocytes (×10^3/ml)</td>
<td>8.7 ± 1.7</td>
<td>9.1 ± 1.0 (NS)</td>
<td>10.7 ± 1.5</td>
</tr>
<tr>
<td>Neutrophils (×10^3/ml)</td>
<td>1.4 ± 0.4</td>
<td>1.5 ± 0.3 (NS)</td>
<td>1.7 ± 0.5 (NS)</td>
</tr>
<tr>
<td>Basophils (×10^3/ml)</td>
<td>0.10 ± 0.04</td>
<td>0.12 ± 0.02 (NS)</td>
<td>0.12 ± 0.03 (NS)</td>
</tr>
<tr>
<td>Monocytes (×10^3/ml)</td>
<td>0.15 ± 0.06</td>
<td>0.15 ± 0.04 (NS)</td>
<td>0.18 ± 0.05 (NS)</td>
</tr>
<tr>
<td>Eosinophils (×10^3/ml)</td>
<td>0.19 ± 0.07</td>
<td>0.17 ± 0.08 (NS)</td>
<td>0.16 ± 0.08 (NS)</td>
</tr>
</tbody>
</table>

Determined with an Advia hematology analyzer (n = 17). Unpaired 2-tailed Student’s t test with Bonferroni correction for multiple comparisons. *P < 0.01. HCT, hematocrit.

### Table 3. Analysis of blood progenitor populations in the bone marrow of adult Tpm4^+/+ and Tpm4^Plt53/− mice by flow cytometry (n = 3)

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Marker</th>
<th>+/-</th>
<th>+/-/Plt53</th>
</tr>
</thead>
<tbody>
<tr>
<td>LSK (+10^6)</td>
<td>LinKit+ sca1+</td>
<td>1.6 ± 0.3</td>
<td>2.2 ± 0.5 (NS)</td>
</tr>
<tr>
<td>MEP (+10^6)</td>
<td>LinKit+ sca1+ CD34+CD16/32a</td>
<td>6.6 ± 0.8</td>
<td>6.3 ± 1.3 (NS)</td>
</tr>
<tr>
<td>MKP (+10^6)</td>
<td>LinKit+ sca1+CD65+CD41+</td>
<td>1.8 ± 0.3</td>
<td>1.8 ± 0.7 (NS)</td>
</tr>
<tr>
<td>CMP (+10^6)</td>
<td>LinKit+ sca1+ CD34+CD16/32a</td>
<td>5.1 ± 0.8</td>
<td>4.8 ± 1.2 (NS)</td>
</tr>
<tr>
<td>GMP (+10^6)</td>
<td>LinKit+ sca1+CD34+CD16/32a</td>
<td>17.3 ± 1.9</td>
<td>18.6 ± 4.4 (NS)</td>
</tr>
</tbody>
</table>

Lineage (Lin) markers included CD4, CD8, CD11b, CD19, B220, Gr1, Ter119, F4/80, and CD11b. The absolute cell number is shown. Unpaired 2-tailed Student’s t test. MKP, megakaryocyte progenitors. LSK, lineage-negative (Lin´), Sca-1¨, c-Kit+ cells; MEP, megakaryocyte–erythroid progenitor; CMP, common myeloid progenitor; CMP, granulocyte–macrophage progenitor.
TPM4 localizes to proplatelets in human and mouse megakaryocytes. Sorting of tropomyosin isoforms to different cellular compartments is thought to be crucial for their function (9). TPM4 localized to proplatelets and colocalized with filamentous (F) actin in primary CD34+-derived human megakaryocytes (Figure 7A). In mature, WT mouse megakaryocytes, TPM4 showed a diffuse, mainly peripheral distribution and did not obviously colocalize with F-actin (Figure 7B). As expected, we could not detect TPM4 in Tpm4Plt53/Plt53 megakaryocytes using the δ/9d antibody (Supplemental Figure 6A). The distribution and morphology of F-actin and tubulin were similar in mature WT and Tpm4Plt53/Plt53 megakaryocytes (not shown). In WT megakaryocytes commencing proplatelet formation, TPM4 was organized in a fine punctuate network (Supplemental Figure 6B). TPM4 colocalized with actin in proplatelet swellings and tips and partially at proplatelet membranes (Figure 7C).

TPM4 facilitates proplatelet formation in human and mouse. We next examined the effect of depleting TPM4 in human megakaryocytes derived from CD34+ hematopoietic progenitor cells. While megakaryocyte proliferation and maturation were unaffected by transduction with shRNAs targeting TPM4 (Supplemental Figure 7A), the level of proplatelet formation was reduced in comparison with the nontargeting control. ShRNA_TPM4B and shRNA_TPM4E, which knocked down mRNA levels to approximately 20% and 8%, respectively, reduced proplatelet formation by approximately 40% and 14%. Logistic regression analysis showed that the correlation between the proportion of proplatelet-forming cells and RNA and protein levels for TPM4 was highly significant (P < 2×10^-16 and < 6.23×10^-12, respectively) (Figure 8A), demonstrating that the level of TPM4 is rate-limiting for platelet formation.

In mice, we observed a significant increase in megakaryocyte numbers in Tpm4Plt53/+ and Tpm4Plt53/Plt53 bone marrow (Figure 8B). Ploidy profiles were unchanged (Supplemental Figure 7B). However, mutant megakaryocytes were smaller than WT counterparts (~20% of Tpm4Plt53/+ and ~30% of Tpm4Plt53/Plt53 compared with only ~4% of Tpm4+/+ megakaryocytes were smaller than 50 μm²), and many exhibited an irregular morphology (Figure 8, C and D). Relative to the WT, proplatelet formation was reduced by approximately 40% in fetal liver–derived Tpm4Plt53/Plt53 megakaryocytes and by approximately 20% in Tpm4Plt53/Plt53 cells (Figure 8E). Time-lapse video microscopy indicated that, while the initial morphological changes associated with proplatelet formation took place in mutant cells, loss of TPM4 resulted in megakaryocytes with few or no branches (Figure 8F and Supplemental Videos 1 and 2). Where proplatelets did form, their tips were significantly increased in size relative to WT counterparts (Supplemental Figure 7C). Visualization of F-actin and tubulin demonstrated that within the short, thick branches produced by Tpm4+/+ megakaryocytes, massive
nia, and have been linked to the RHOA/ROCK pathway (33). We therefore examined the expression and phosphorylation state of RHOA, ROCK, and NMMHC-IIa by Western blot. No differences between Tpm4Plt53/Plt53, Tpm4Plt53/+ and Tpm4+/+ megakaryocytes and platelets were observed (Supplemental Figure 10A). Confocal microscopy of Tpm4+/+ megakaryocytes undergoing the first morphological changes presaging proplatelet formation revealed that NMMHC-IIa localized to a fine network (Supplemental Figure 10B), very similar to the distribution of TPM4 (Supplemental Figure 6B). This pattern was dramatically perturbed in approximately 50% of Tpm4Plt53/Plt53 megakaryocytes. In WT proplatelet swellings and tips, NMMHC-IIa localized to a previously described characteristic network and to membranes (Figure 9C, left, and ref. 34). In contrast, NMMHC-IIa in Tpm4Plt53/Plt53 proplatelet-forming megakaryocytes showed a patchy distribution in approximately 70% of cells analyzed, as compared with approximately 15% of Tpm4+/+ counterparts (Figure 9C, right). These results indicate that TPM4 deficiency impacts multiple regulators of the megakaryocyte actin cytoskeleton.

Discussion

Cytoskeletal reorganizations are the driving force for the production of platelets by megakaryocytes (35). We therefore examined the expression and phosphorylation state of RHOA, ROCK, and NMMHC-IIa by Western blot. No differences between Tpm4+/+ mice and Tpm4Plt53/Plt53 megakaryocytes and platelets were observed (Supplemental Figure 10A). Confocal microscopy of Tpm4+/+ megakaryocytes undergoing the first morphological changes presaging proplatelet formation revealed that NMMHC-IIa localized to a fine network (Supplemental Figure 10B), very similar to the distribution of Tpm4+/+ megakaryocytes. This pattern was dramatically perturbed in approximately 50% of Tpm4Plt53/Plt53 megakaryocytes. In WT proplatelet swellings and tips, NMMHC-IIa localized to a previously described characteristic network and to membranes (Figure 9C, left, and ref. 34). In contrast, NMMHC-IIa in Tpm4+/+ proplatelet-forming megakaryocytes showed a patchy distribution in approximately 70% of cells analyzed, as compared with approximately 15% of Tpm4+/+ counterparts (Figure 9C, right). These results indicate that TPM4 deficiency impacts multiple regulators of the megakaryocyte actin cytoskeleton.

Figure 5. The function of human platelets is mildly affected by reduced TPM4 expression. (A) Platelet function testing measuring fibrinogen binding (top) and P-selectin expression (bottom) (percentage of positive platelets) after treatment with final concentrations of 0.0005 μM ADP, 0.3 μg/ml CRP-XL, or 0.8 μM TRAP-6. Depicted are the results obtained on 3 different days (2 technical replicates) for the day control and the patient (case 1-III-7). A bank of 20 controls is shown in white. (B) and (C) Thrombus formation on collagen under flow (shear rate 1,600/s). (B) Normalized thrombus number and coverage after blood perfusion through a collagen-coated chamber; 6 images captured per run, 2 runs per sample using 2 different samples. (C) Representative images of thrombi stained with P-selectin captured using fluorescence microscopy (EVOS system; Advanced Microscopy Group). Scale bars: 50 μm. Unpaired 2-tailed Student’s t test. ***P < 0.001.
Figure 6. The function of mouse platelets is mildly affected by reduced TPM4 expression. (A) Normal in vivo lifespan of Tpm4<sup>+/+</sup> and Tpm4<sup>Plt53/Plt53</sup> compared with Tpm4<sup>−/−</sup> platelets. Platelets were labeled by i.v. injection with X-488 antibody (Emfret), and the percentage of fluorescent platelets was monitored over time by flow cytometry (n = 5, representative of 2 independent experiments). (B) Flow cytometric measurement of integrin α<sub>i</sub>β<sub>3</sub> activation (JON/A–PE antibody) in Tpm4<sup>−/−</sup>, Tpm4<sup>Plt53/Plt53</sup>, and Tpm4<sup>Plt53/Plt53</sup> platelets after activation with the depicted agonists (n = 4, representative of 3 independent experiments). (C) Platelet spreading. Left: Representative differential interference microscopy images of Tpm4<sup>−/−</sup> and Tpm4<sup>Plt53/Plt53</sup> platelets spread on fibrinogen (100 μg/ml) after activation with 0.01 U/ml thrombin. Scale bar: 5 μm. Right: Percentage of fully spread Tpm4<sup>−/−</sup> (black) and Tpm4<sup>Plt53/Plt53</sup> (blue) platelets at 15, 30, and 60 minutes after induction of the spreading process (n = 3). Results are representative of 3 independent experiments. (D) Thrombus formation on collagen under flow (shear rate 1,000/s). Left: Mean surface covered with thrombi. Right: Relative platelet deposition, as measured by integrated fluorescent intensity per square millimeter ± SD. Each dot represents an individual. Results are pooled from 2 independent experiments. (E) Tail bleeding times in Tpm4<sup>Plt53/Plt53</sup> (blue) and Tpm4<sup>Plt53/Plt53</sup> (light blue) compared with Tpm4<sup>−/−</sup> (black) mice. Each dot represents an individual. Unpaired 2-tailed Student’s t test, *P < 0.05, **P < 0.01. Our results indicate that TPM4 insufficiency affects multiple cytoskeletal regulators with known roles in platelet production, including cofillin, myosin IIa, actinin 1, and filamin A. The abnormal distribution of tubulin, polymerization of which is considered a driving force for proplatelet elongation (2, 40), was potentially a downstream effect. Despite recent work suggesting that capping of TPM4-bound actin filaments by TMOD3 is required for proplatelet formation (30), TMOD3 localization was not obviously affected in Tpm4 mutant platelets. This might be explained by TPM1 or TPM3.1 partially compensating for the lack of TPM4 during proplatelet formation.

Mice lacking cofillin in the megakaryocyte lineage display macrothrombocytopenia (41), suggesting that the correct balance between TPM4 and cofillin is essential for the production of correctly sized platelets. The decreased levels of inactive, phosphorylated, cofillin in Tpm4 mutant megakaryocytes and platelets are consistent with previous studies demonstrating that tropomyosin and cofillin compete with each other at the actin filament (29), and indicate that reduced TPM4 expression facilitates increased binding of active cofillin to actin filaments. Mutations in the gene encoding NMMHC-IIa (MYH9) in cases with MYH9-related disorders cause autosomal dominant macrothrombocytopenia and bleeding associated with other complex pathobiologies (42). Altered localization of NMMHC-IIa was observed in a large proportion of Tpm4 mutant megakaryocytes. This finding is in accordance with a previously established role for TPM4.2 in myosin II recruitment to stress fiber precursors (31) and its specific regulatory impact on NMMHC-IIa (43).

Reduced TPM4 expression resulted in the production of large platelets with fragile appearance (Figure 4 and Supplemental Figure 4). Fragile platelets with impaired function are a characteristic of patients with FLNA mutations (24, 44), and of Flna-null mice (45). We observed increased levels of degraded filamin A in Tpm4 mutant platelets. Interaction of filamin A with glycoprotein (GP1B), a component of the VWF receptor complex, is important for regulation of platelet size, as well as platelet adhesion and maintenance of membrane integrity under high shear (46–48). Thus, disturbances in interactions between filamin A and F-actin may contribute to the altered size and function of TPM4-bound actin filament arrangements in megakaryocytes and platelets. Our findings therefore suggest that the amount of available TPM4-bound actin filaments in megakaryocytes directly determines the amount of membrane regions, which can generate proplatelet branches.
deficient platelets. Tpm4 mutant platelets also exhibited partial degradation of actinin 1. Mutations in the latter cause autosomal dominant thrombocytopenia (25–27). Prior studies have suggested that the levels of actin-binding proteins correlate with fluctuations in tropomyosin isoforms (36, 49). Our work indicates that loss of TPM4 binding inhibits binding of other proteins, such as filamin A and actinin 1, to actin filaments. This results in an impairment of the complex actomyosin cytoskeletal network that drives proplatelet formation.

TPM4 insufficiency affected neither the in vivo platelet lifespan in mice, nor the in vitro platelet functional responses to agonists in humans and mice. The delayed spreading observed in Tpm4 mutant platelets may be attributed to their increased size, but may also indicate specific defects upon decreased TPM4 expression, potentially involving altered integrin αIIbβ3 outside-in signaling (Figure 7C). Consistent with this, mutant mice, and WT mice with transplanted mutant bone marrow, tended toward prolonged tail bleeding times. Furthermore, thrombus formation on collagen under flow by TPM4-deficient platelets of both human and mouse origin was reduced. Since the platelet disorder associated with the TPM4 variant results in an unpredictable bleeding phenotype, these findings indicate that reduced levels of TPM4 result in a mild impairment of hemostasis.

Our findings indicate that, like the targeted knockout, the TPM4 variants we identified in both humans and mice result in a complete loss of function at the TPM4 locus. The fact that mice homozygous for the Plt53 mutation or null allele are healthy and fertile was very surprising, standing in stark contrast to mice lacking Tpm1, Tpm2, or Tpm3, which die in utero (11–13). Thus, TPM4 is the only tropomyosin family member dispensable for development. It plays an essential and highly specific role in F-actin organization in the megakaryocyte lineage. Given the exceptionally high level of conservation between them, the inability of other tropomyosin isoforms to fully compensate for the loss of TPM4 deletion highlights a profound gap in our understanding of how functional specificity is achieved and governed.

Methods

Mice

Generation of mice with a mutant Tpm4 allele. Mice carrying a mutant Tpm4 allele, named Plt53, were generated. Briefly, male BALB/c mice were injected i.p. with 85 mg/kg ENU weekly for 3 weeks (50). Treated animals were rested for 12 weeks before mating with untreated BALB/c females to produce first-generation progeny. Peripheral blood was taken from these mice at 7 weeks of age and a complete blood cell count performed using an Advia 2120 hematological analyzer. Animals exhibiting aberrant platelet counts were test-mated to determine heritability of the phenotype. Mutant strains were backcrossed for 10 generations to the BALB/c background in order to breed out irrelevant ENU-induced mutations. Tpm4mut/mut mice were intercrossed to yield Tpm4mut/+, Tpm4mut/mut, and Tpm4mut/mut Intermates, which were used in experiments. Mice were 7 to 12 weeks old or as otherwise stated.

Figure 7. TPM4 localizes to proplatelets in human and mouse megakaryocytes. (A) Immunolabeling of TPM4 (red), F-actin (purple), α-tubulin (green), and merge of TPM4/F-actin/α-tubulin and DAPI, in control human proplatelets. Scale bar: 25 μm. (B) Investigation of TPM4 (yellow) and F-actin (red) localization of mouse fetal liver cell–derived megakaryocytes by confocal immunofluorescence microscopy. TPM4 (yellow) localizes to the periphery in mature round WT Tpm4+/+ megakaryocytes. (C) TPM4 is enriched in proplatelet tips and colocalizes with F-actin (depicted by arrows). Scale bars: 20 μm.
Figure 8. TPM4 dose-dependently facilitates proplatelet formation. (A) Logistic regression analysis showing that the percentage of proplatelet-forming cells decreases with Tpm4 RNA (left; effect 1.22, \( P < 2^{-16} \)) and protein (right; effect 2.54, \( P = 6.23^{-12} \)) levels. Node size reflects the number of cells counted (at least 200). (B) Increased number of megakaryocytes (MK) in bone marrow from Tpm4\(^{-/-}\) and Tpm4\(^{-/-}\) and 28 FOV (Tpm4\(^{+/-}\)). (C) Representative pictures of H&E-stained bone marrow sections show altered morphology (smaller size, irregular shape) of Tpm4\(^{-/-}\) and Tpm4\(^{+/+}\) megakaryocytes compared with the control (\( n = 3 \)). Arrows indicate megakaryocytes. Scale bar: 15 \( \mu \)m. (D) Decreased size of Tpm4 mutant megakaryocytes compared with WT counterparts. \( n = 105 \) (Tpm4\(^+/+\)), \( n = 35 \) (Tpm4\(^{-/-}\)), \( n = 125 \) (Tpm4\(^{+/+}\)). (E) Left: Decreased proplatelet formation (PPF) of fetal liver cell-derived Tpm4\(^{-/-}\) and Tpm4\(^{+/+}\) megakaryocytes (\( n = 5-6 \)). Each dot represents the mean of 1 individual sample (at least 12 visual fields counted). Right: Representative light microscopy pictures showing altered morphology and decreased branch formation of Tpm4\(^{-/-}\) and Tpm4\(^{+/+}\) megakaryocytes. Scale bar: 50 \( \mu \)m. (F) Investigation of F-actin (red) and tubulin (green) distribution of proplatelet-forming Tpm4\(^{+/+}\) (left) and Tpm4\(^{+/+}\) (right) megakaryocytes by confocal immunofluorescence microscopy. Nuclei were stained with DAPI (blue). Scale bars: 20 \( \mu \)m. **P < 0.01, ***P < 0.001.
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The enumeration of hematopoietic progenitors in mice by flow cytometry

Organ cellularity was determined by counting of cell suspensions from bone marrow, spleen, and thymus with calibrated beads (BD Biosciences). Cells were stained on ice for 30 minutes, washed once, and then incubated with streptavidin–PE–Texas red (Life Technologies). After a washing step, acquisition was performed on an LSR Fortessa (BD Biosciences) and analyzed with FlowJo (Tree Star).

Antibodies for flow cytometry either were labeled in-house with FITC, PE, PECy7, APC, or Alexa Fluor 700 or were biotinylated. CD4 (GK1.5), CD8 (53.6.7), CD16/32 (24G2), CD19 (1D3), CD41 (MWReg30), Ter119 (Ly76), Gr1 (RB6-8C5), CD11b (M1/70), Sca1 (D7), Kit (ACK4), B220 (RA3-6B2), and CD34-FITC (RAM34) were purchased from BD Pharmingen, and CD150-PE (TC15-12F12.2) was purchased from BioLegend. Fluorogold (Sigma-Aldrich) was used to exclude dead cells.

Generation of Tpm4.2 knockout mice. Target sites in exon 1b of the mouse Tpm4 gene were analyzed by the MIT CRISPR Design Tool (51). Two high-scoring (>85) guide RNA (gRNA) sequences were chosen, as they created a 93-bp deletion that interrupted the ATG start codon. The mouse oocytes were preincubated in cytochalasin B (5 μg/ml); then 12.5 ng/μl of each commercially synthesized gRNA (Sigma-Aldrich) and 50 ng/μl of Cas9 mRNA were coinjected into the cytoplasm of mouse oocytes (52). Progeny were screened for the 93-bp deletion using the primers Tpm4.2 total KO forward 5′-GTGACCTCATGGGCCTGAC-3′ and Tpm4.2 total KO reverse 5′-GGACGAAAGTGGAATCGG-3′, which detects a WT allele of 317 bp and a knockout allele of 223 bp. DNA from founder mouse 6 had the expected 90-bp deletion, and a subsequent Western blot of tail lysates revealed the complete absence of TPM4.2, the sole TPM4 protein isoform expressed in mice.

Hematology

Human. Automated full blood cell counts were performed on blood collected by venipuncture into tubes containing EDTA using a Sysmex XN-1000 or XE-5000 hematological analyzer.

Mouse. Automated cell counts were performed on blood collected by cardiac puncture or from the retro-orbital plexus into Microtainer tubes containing EDTA (Sarstedt), using an Advia 2120 hematological analyzer (Siemens). Megakaryocytes were counted manually in sections of sternum stained with H&E with a minimum of 10 visual fields (×200) analyzed. Images were acquired on a Nikon Eclipse E600 microscope equipped with AxioCam MRc5 (Zeiss) and AxioVision 4.8. Scale bars were inserted and the megakaryocyte area was determined using ImageJ (NIH). Acute thrombocytopenia was induced by injection of antiplatelet serum and assessed as previously described (53).

Enumeration of hematopoietic progenitors in mice by flow cytometry

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Investigation of the effect of the Plt53 mutation in megakaryocytes by RT-PCR

Fetal liver cell–derived megakaryocytes from Tpm4+/+, Tpm4Plt53/+, and Tpm4Plt53/Plt53 embryos E13.5 were generated as described below. mRNA was isolated using the RNeasy Plus Kit (Qiagen), and cDNA was synthesized using SuperScript III Reverse Transcriptase (Invitrogen) according to the manufacturer’s protocols. The primers used were 5′-AAAGAAGCCAAGCACATCAC-3′ (Tm4_F3) and 5′-AGC-CCACATTCTCTTCTTTG-3′ (Tm4_R7). The expected product size

Figure 9. TPM4-interacting proteins in megakaryocytes and platelets. (A) Investigation of cofilin and P-cofilin expression in Tpm4+/+, Tpm4Plt53/+, and Tpm4Plt53/Plt53 compared with Tpm4+/+ platelets (n = 6; results were pooled from 2 separate experiments). (B) Densitometry analysis shows decreased levels of phosphorylated (inactive) cofilin in Tpm4+/+, Tpm4Plt53/+, and Tpm4Plt53/Plt53 platelets (n = 6; results were pooled from 2 separate experiments). (C) Investigation of NMMHC-Ⅱa (green) and F-actin (red) localization in fetal liver cell–derived Tpm4+/+ and Tpm4Plt53/Plt53 megakaryocytes by confocal immunofluorescence microscopy. Scale bar: 20 μm. One-way ANOVA, unpaired 2-tailed Student’s t test with Bonferroni correction for multiple comparisons. *P < 0.05.
for the WT was 360 bp, while an approximately 400-bp product was obtained in the presence of the Tpm4Δ allele. PCR products were sequenced and revealed the presence of an approximately 40-bp intronic inclusion in the mutant compared with the WT product.

**Gene expression analysis in human megakaryocytes by RT-quantitative PCR**

Total RNA was extracted using an RNasy Plus Kit (Qiagen) according to the manufacturer’s instructions. cDNA was prepared from 250–500 ng total RNA using SuperScript III First-Strand Synthesis system for RT-PCR (Invitrogen) with random hexamers. Two-step quantitative PCR (qPCR) reactions were performed in duplicates using SYBR Green chemistry (Brilliant II SYBR Green QPCR Low ROX, Agilent Technologies) on an Mx3000P instrument (Agilent Technologies). Relative gene expression was calculated by the ΔΔCt method using GAPDH housekeeping gene for normalization. PCR primer pairs were designed to amplify only cDNA, had no reported off-target matches searching the human NCBI RefSeq database, and were within 80%–120% PCR efficiencies with single dissociation curves.

qPCR oligonucleotide primer pairs were as follows: TPM4-F, AGTCGAGCCCAAAAAAGATGA; TPM4-R, TCGCTCTTAAATGATGA; TPM3-F, GGAGATGAGGTGGATACTCGAGTAGTATCCACGACTAGATACTGGGTTTTTG; TPM4_shE reverse, CCGGCCCAGTATCTAGTCGTGGATACTCGAGTTAAGTTCGTTTAGTGTCTGA; TPM4_shB reverse, AATTCAAAAATCAGACACTACAACTCGAGTTAAGTTCGTTTAGTGTCTGA; TPM4_shB forward, CCGGTCAGACACTACAACTCGAGTTAAGTTCGTTTAGTGTCTGA; TPM4_shA reverse, AATTCAAAAATCAGACACTACAACTCGAGTTAAGTTCGTTTAGTGTCTGA; TPM4_shA forward, CCGGGGAAGAGGCTGACCGCAAATACTCGAGTATTGAGTATCCACGACTAGATACTGGGTTTTTG; TPM4_shE reverse, AATTCAAAAATCAGACACTACAACTCGAGTTAAGTTCGTTTAGTGTCTGA; TPM4_shE forward, CCGGTCAGACACTACAACTCGAGTTAAGTTCGTTTAGTGTCTGA.

**TPM4 knockdown in human megakaryocytes**

**Short hairpin vector cloning.** Oligonucleotides were selected from the RNAi consortium database. Short hairpin or control nontarget sequences were annealed before cloning into the pLKO.1 lentiviral vector backbone (Addgene 10878) downstream of the human U6 promoter and checked for sequence integrity.

**Viral particle production.** Replication-deficient lentiviral vector particles (LVPs) were produced by transient cotransfection of HEK 293T/17 cells (ATCC CRL-11268) with pLKO.1–short hairpin constructs along with the psPAX2 and pMD2.G helper plasmids (Addgene 12260, 12259) using TurboFect transfection reagent (Thermo Fisher Scientific). Crude supernatants containing LVPs were concentrated by centrifugation at 1,000 g for 8 minutes, followed by centrifugation of the supernatant Buffy coat at 125 g for 8 minutes. Platelets were washed by 2 sequential centrifugations at 860 g for 5 minutes in 140 mM NaCl, 5 mM KCl, 12 mM trisodium citrate, 10 mM glucose, and 12.5 mM sucrose, pH 6.0. The platelet pellet was resuspended in Tyrode-HEPES buffer (134 mM NaCl, 0.34 mM NaHPO4, 2.9 mM KCl, 12 mM NaHCO3, 5 mM HEPES, pH 7.4) containing 5 mM glucose, 0.35% BSA, and 1 mM CaCl2.

**Analysis of platelet activation and surface receptor expression by flow cytometry**

**Mouse.** Washed platelets were adjusted to a concentration of 5 × 10⁴ per microliter and activated with the indicated agonists for 15 minutes at room temperature in the presence of JON/A-PE and anti-P-selectin–FITC antibodies (diluted according to the manufacturers’ instructions). Samples were diluted with PBS and analyzed on a FACScalibur. To assess platelet receptor expression, anticoagulated whole blood was diluted 1:20 using Tyrode-HEPES buffer and stained with platelet-specific antibodies or negative controls for 20 minutes at room temperature. Samples were diluted with Tyrode-HEPES buffer and directly acquired by flow cytometry.

**Human.** Citrated whole blood was incubated for 5 minutes with hirudin and aspirin, before being added to tubes containing HEPES-buffered saline, FITC–rabbit anti-fibrinogen antibody (Dako, F0111), and PE–mouse anti-P-selectin (IBGRL, clone Thromb6) or relevant negative controls (P-selectin isotype, IBGRL, clone 9E10). Platelets were agonized with a final concentration of 0.0005 μM ADP, 0.3 μg/ml CRP-XL, and 0.8 μM TRAP-6 or left untreated. Apyrase was added to platelets agonized with CRP-XL and TRAP-6. Flow cytometry was performed on an FC-500 (Beckman Coulter), and platelets were identified by light scatter. Results were recorded as the percentage of platelets positive for the relevant activation marker.

**Platelet spreading**

Coverslips were coated with 100 μg/ml human fibrinogen and blocked with PBS 1% BSA. After rinsing with Tyrode-HEPES buffer, washed mouse platelets (100 μl with 0.03 × 10⁵ platelets per microliter) were activated with 0.005 U/ml thrombin and incubated at room temperature.
tute for the indicated time periods. After fixation with 4% paraformaldehyde in PBS for 10 minutes, the coverslips were thoroughly rinsed with PBS, and platelets were visualized with a DeltaVision Elite microscope equipped with CoolSnap2 CCD Detector (>100/1.4 oil immersion objective). Images were analyzed using ImageJ software.

Platelet turnover studies in mice
Mice were injected i.v. with 0.15 μg/g body weight of X-488 (Emfret), a rat-derived IgG against the platelet glycoprotein Ibα receptor, and platelet lifespan was measured as previously described (54).

Tail bleeding assay
Mice were anesthetized using ketamine/xylazine. The tail was transected at 2 mm from the tip and immediately immersed in 37°C saline. The bleeding time was determined as the time from the tail transection to the cessation of blood flow or stopped after 10 minutes.

Statistics
Data distribution was analyzed using the Shapiro-Wilk test. Statistical significance between 2 experimental groups was analyzed using an unpaired 2-tailed Student’s t test or a Mann-Whitney test. When more than 2 experimental groups were compared, data were analyzed using 1-way ANOVA. When ANOVA indicated that a significant difference was present, individual differences were explored with the 2-tailed unpaired Student’s t test or Mann-Whitney test using Bonferroni correction for multiple comparisons (Prism 7, GraphPad Software). P values less than 0.05 were considered as statistically significant; *P < 0.05, **P < 0.01, ***P < 0.001 or as otherwise stated. Data are presented as mean ± SD.

Study approval
Patients. The patients were enrolled to the BRIDGE-BPD study (UK REC10/H0304/66) and to the NIHR BioResource–Rare Diseases study (UK REC 13/EE/0325) after providing informed written consent. The control volunteers for functional work were enrolled to the Genes and Platelets study (UK REC10/H0304/65) after providing informed consent. For genetic analysis, control groups comprised participants was supported by the NIHR BioResource–Rare Diseases project of the NIHR BioResource–Rare Diseases. PWG contributed to the design and interpretation of experiments on the detection and function of tropomyosins. ET and KD, and WHO). The NIHR BioResource–Rare Diseases is responsible for the delivery of the rare diseases pilot phase of the 100,000 Genomes Project and is funded by the National Institute for Health Research (NIHR; http://www.nihr.ac.uk). Research in the Ouwehand laboratory also receives funding support from the European Commission, NIHR, Wellcome Trust, Medical Research Council (MRC), and British Heart Foundation under numbers RP-PG-0310-1002 and RG/09/12/28096. We are grateful to Janelle E. Collinge for outstanding technical assistance with gene mapping studies. We thank E. Major, S. Ross, T. Carle, T. Gibbs, and S. Green for outstanding animal husbandry assistance, J. Corbin for Advia analysis, and Emma C. Joseffson for the purification of human platelets. Confocal microscopy work on human samples was supported by the Cambridge NIHR Biomedical Research Centre Cell Phenootyping Hub. We thank Jeremy Skepper and Janet Powell of the Cambridge University Advanced Imaging Centre for technical support with electron microscopy imaging. SWK is supported by an MRC Clinical Training Fellowship (MR/K023489/1). ADM receives support from the Bristol NIHR Biomedical Research Unit for Cardiovascular Disease. This work was supported by a Project Grant (no. 575535), a Program Grant (no. 1016647), a Fellowship (1063008 to BTK and 1058344 to WSA), Project Grants (to PWG...
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Address correspondence to: Benjamin T. Kile, ACRF Chemical Biology Division, Walter and Eliza Hall Institute of Medical Research, 1G Royal Parade, Parkville, Victoria 3052, Australia. Phone: 61.3.9345.2555; E-mail: kile@wehi.edu.au. Or to: Marloes R. Tijssen, Department of Haematology, University of Cambridge, Long Road, Cambridge CB2 0PT, United Kingdom. Phone: 441223.588166; E-mail: mrt37@cam.ac.uk.

IP’s present address is: Institute of Experimental Biomedicine, University Hospital and Rudolf Virchow University, Würzburg, Würzburg, Germany.