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Chronic inflammation in visceral adipose tissue (VAT) precipitates the development of cardiometabolic disorders. Although changes in T cell function associated with visceral obesity are thought to affect chronic VAT inflammation, the specific features of these changes remain elusive. Here, we have determined that a high-fat diet (HFD) caused a preferential increase and accumulation of CD44$^{hi}$CD62L$^{lo}$CD4$^+$ T cells that constitutively express PD-1 and CD153 in a B cell–dependent manner in VAT. These cells possessed characteristics of cellular senescence and showed a strong activation of Spp1 (encoding osteopontin [OPN]) in VAT. Upon T cell receptor stimulation, these T cells also produced large amounts of OPN in a PD-1–resistant manner in vitro. The features of CD153$^+$PD-1$^+$CD44$^{hi}$CD4$^+$ T cells were highly reminiscent of senescence-associated CD4$^+$ T cells that normally increase with age. Adoptive transfer of CD153$^+$PD-1$^+$CD44$^{hi}$CD4$^+$ T cells from HFD-fed WT, but not Spp1-deficient, mice into the VAT of lean mice fed a normal diet recapitulated the essential features of VAT inflammation and insulin resistance. Our results demonstrate that a distinct CD153$^+$PD-1$^+$CD44$^{hi}$CD4$^+$ T cell population that accumulates in the VAT of HFD-fed obese mice causes VAT inflammation by producing large amounts of OPN. This finding suggests a link between visceral adiposity and immune aging.

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Chronic inflammation in visceral adipose tissue (VAT) precipitates the development of cardiometabolic disorders. Although changes in T cell function associated with visceral obesity are thought to affect chronic VAT inflammation, the specific features of these changes remain elusive. Here, we have determined that a high-fat diet (HFD) caused a preferential increase and accumulation of CD4+CD62L−CD4+ T cells that constitutively express PD-1 and CD153 in a B cell–dependent manner in VAT. These cells possessed characteristics of cellular senescence and showed a strong activation of Spp1 (encoding osteopontin [OPN]) in VAT. Upon T cell receptor stimulation, these T cells also produced large amounts of OPN in a PD-1–resistant manner in vitro. The features of CD153−PD-1−CD44−CD4+ T cells were highly reminiscent of senescence-associated CD4+ T cells that normally increase with age. Adoptive transfer of CD153−PD-1−CD44−CD4+ T cells from HFD-fed WT, but not Spp1-deficient, mice into the VAT of lean mice fed a normal diet recapitulated the essential features of VAT inflammation and insulin resistance. Our results demonstrate that a distinct CD153−PD-1−CD44−CD4+ T cell population that accumulates in the VAT of HFD-fed obese mice causes VAT inflammation by producing large amounts of OPN. This finding suggests a link between visceral adiposity and immune aging.

Introduction

Visceral obesity is associated with chronic low-grade inflammation in visceral adipose tissue (VAT) and a sustained whole-body proinflammatory state, which may underlie metabolic and cardiovascular diseases (1–6). VAT inflammation associated with obesity involves a complex network of responses of immune cell components, including acquired immune cells such as various subsets of T cells and B cells and innate immune cells such as macrophages (7–10). Among these cells, CD4+ T cells have been recognized as a central regulator of chronic VAT inflammation (9–14). The number of CD4+ T cells in VAT increases as the tissue expands in obesity. Factors that drive CD4+ T cell expansion and differentiation into proinflammatory effectors in VAT during the development of high-fat diet–induced (HFD-induced) obesity may include MHC class II–associated antigens, possibly self-peptides, because the T cell receptor (TCR) repertoire of CD4+ T cells in VAT is limited, and deficiency of MHC class II protects mice from HFD-induced VAT inflammation and insulin resistance (11, 14–16). For instance, IFN-γ–producing Th1 cells enhance proinflammatory macrophage activation (12–14, 17), and IL-17 produced by Th17 cells may cause insulin resistance by affecting insulin receptor signaling (18). By contrast, IL-4– and IL-13–secreting Th2 cells and FoxP3+ Tregs suppress VAT inflammation by inducing antiinflammatory macrophages that secrete IL-10 (19–21). However, the obesity-associated immune background underlying chronic inflammation in VAT remains elusive.

Significant changes occur in the overall T cell populations with age. In CD4+ T cells, proportions of naive (CD44hiCD62Lhi) cells sharply decline in ontogeny, with an age-dependent increase in cells of the memory phenotype (CD44hiCD62Llo) (22, 23). Among CD44hiCD4+ T cells, a unique population expressing programmed cell death 1 (PD-1) and CD153 actually increases with age in mice (20). PD-1 is a negative costimulatory receptor for TCR signaling (24), and CD153 is a TNF superfamily protein (25). The CD153−PD-1−CD44−CD4+ T cell population shows compromised proliferation and regular T cell cytokine production on T cell receptor (TCR) stimulation but secretes large amounts of proinflammatory cytokines, such as osteopontin (OPN). These CD4+ T cells also show signatures of cell senescence, including a marked increase in senescence-related gene expression and nuclear heterochromatin foci, and are termed senescence-associated T cells (SA–T cells) (9). Notably, the age-dependent development of SA–T cells, which may include autoreactive cells, is dependent on B cells (9). As such, the increase in SA–T cells is suggested to be involved in part in immune aging phenotypes such as impaired acquired...
immune capacity, increased proinflammatory traits, and high risk for autoimmunity (26).

In the present study, we demonstrate that CD153+PD-1+ CD44hiCD4+ T cells are remarkably increased and preferentially accumulated in the VAT of HFD-fed mice in a B cell–dependent manner and that these CD4+ T cells show functional and genetic features strongly resembling SA-T cells that increase in secondary lymphoid tissues with age. We also indicate that the CD153+PD-1+CD44hi CD4+ T cells play a crucial role in inducing chronic VAT inflammation and metabolic disorder via secretion of large amounts of OPN.

Results

A HFD induces an increase and accumulation of PD-1+CD44hiCD62Llo CD4+ T cells in VAT. C57BL/6 (B6) mice were fed an HFD starting at 4 weeks of age. By 18 weeks of age, these mice had an increased BW and visceral fat mass, glucose intolerance, and insulin resistance compared with age-matched B6 mice fed a normal diet (ND) (Supplemental Figure 1, A and B; supplemental material available online with this article; doi:10.1172/JCI88606DS1). HFD-fed mice showed remarkably increased numbers of F4/80hiCD11b+ macrophages per gram of VAT, with a drastic shift from a CD11clo CD206hi to a CD11chiCD206lo phenotype (Supplemental Figure 1, C and D). These macrophages were localized in crown-like structures (CLSs) (Supplemental Figure 1E), representing typical obesity-associated chronic inflammation of VAT (17, 27). VAT of 18-week-old HFD-fed obese mice showed increased proportions of CD4+ T cells in total T cell infiltrates (Figure 1A). The absolute numbers of CD4+ T cells per gram of VAT were significantly higher than those in ND-fed mice as early as 2 weeks after initiation of the HFD and progressively increased thereafter (Figure 1A). Notably, the VAT CD4+ T cells from 18-week-old HFD-fed mice showed an increasing shift in the proportions of naive CD44lo CD62Lhi and CD44hiCD62Llo phenotype T cells (Figure 1B), and the increase in absolute numbers of VAT CD4+ T cells was largely attributed to the CD44hiCD62Llo cells (Figure 1B). More than half of the VAT CD44hiCD4+ T cells expressed PD-1, and the numbers of PD-1+CD44hiCD4+ T cells were remarkably higher than those in age-matched ND-fed mice (Figure 2A). Immunostaining analysis confirmed that PD-1+ T cells were localized in CLSs of HFD-fed obese mice, but they were rarely detected in those of ND-fed lean mice (Figures 2B). These results indicate that PD-1+CD44hiCD4+ T cells are predominantly increased in the inflammatory foci of VAT from HFD-fed mice.

Results

The VAT PD-1+CD44hiCD4+ T cell population includes senescent T cells that preferentially produce OPN. We next compared the functional features of PD-1− and PD-1+ CD4+ T cells in VAT from 18-week-old HFD-fed mice. Isolated PD-1−CD44hiCD4+ T cells showed significantly less production of IL-2 and IFN-γ upon TCR stimulation than did PD-1+ CD4+ T cells (Figure 3A), which was consistent with diminished expression of special AT-rich sequence-binding protein 1 (Satb1), a crucial gene for T cell cytokine activation (28, 29), and increased expression of CCAAT/enhancer-binding protein α (Cebpa), which is normally expressed in myeloid cell lineages (30) (Figure 3B). However, the VAT PD-1+CD44hiCD4+ T cells secreted remarkably large amounts of OPN via TCR stimulation, whereas the PD-1− fraction did so minimally (Figure 3C). HFD-fed mice consistently showed significantly increased serum OPN levels (Figure

**Figure 1.** An HFD induces the accumulation of CD44hiCD4+ T cells in VAT. WT mice were fed an HFD for 14 weeks beginning at 4 weeks of age. Age-matched WT mice fed an ND were used as controls. (A) Proportion of VAT CD4+ T cells in total T cells. Temporal dynamics of changes in the number of CD4+ T cells per gram of VAT (n = 5 mice per group). (B) Analysis of VAT CD44hiCD62Llo and CD44hiCD62LloCD4+ T cells (n = 5 mice per group). Data represent the mean ± SEM. *P < 0.05, **P < 0.001, and ***P < 0.0001, by 2-tailed Student’s t test.
weeks of age (Figure 4C). We then isolated 3 distinct populations of CD4+ T cells, PD-1−, CD153−PD-1+, and CD153+PD-1+, from VAT of 18-week-old HFD-fed obese mice and compared their genetic signatures. Expression of secreted phosphoprotein 1 (Spp1), encoding OPN, was markedly high in the CD153+ cells, whereas its expression was minimal in CD153−PD-1+ and PD-1− cells (Figure 5A). CD153+ cells also showed significantly increased expression of cyclin-dependent kinase inhibitor 1A (Cdkn1a, also known as Cip1) and Cdkn2b (also known as Ink4b), which are typical cellular senescence biomarker genes (32), compared with CD153−PD-1+ and PD-1− cells. Both CD153+ and CD153−PD-1+ cells showed downregulation of Satb1, eukaryotic translation elongation factor 1α (Eef1a1), and dual-specificity phosphatase 10 (Dusp10) expression, also suggestive of cellular senescence (33, 34). Furthermore, the vast majority of the PD-1+ cells expressed senescence-associated β-galactosidase (SA–β-gal), a typical cell senescence marker (Figure 3E), and showed remarkably higher expression of γ-H2AX compared with the PD-1− counterpart cells (Figure 3F), which is indicative of greater exposure to genostress. These findings suggest that the CD44hiCD4+ T cell population in VAT of HFD-fed mice includes senescent T cells capable of preferentially producing OPN.

CD153 expression defines a unique PD-1+CD44hiCD4+ T cell population with features of cell senescence that increases almost exclusively in VAT of HFD-fed mice. Because CD153 expression defines the SA–T cells, which gradually increase systemically with age (31), we next examined the expression of CD153 in VAT. We found that a minor population of VAT PD-1+CD4+ T cells in HFD-induced obese mice strongly expressed CD153 (approximately 15% of CD4+ T cells), whereas CD4+ T cells in VAT of ND-fed mice rarely did so (Figure 4A). Other infiltrated cell populations such as CD8+ T cells, macrophages, and B cells in VAT of HFD-fed mice hardly expressed CD153 (Figure 4B). The CD153−PD-1+CD44hiCD4+ T cells were evident in VAT beginning 2 weeks after initiation of the HFD and dramatically increased in mice at 18 weeks of age, thus becoming prominent at later stages in the PD-1+CD44hiCD4+ T cells (Figure 4C; see also Figure 2A). In contrast, this cell population remained negligible in VAT from ND-fed mice at least until 18 weeks of age (Figure 4C). We then isolated 3 distinct populations of CD4+ T cells, PD-1+, CD153−PD-1+, and CD153+PD-1+, from VAT of 18-week-old HFD-fed obese mice and compared their genetic signatures. Expression of secreted phosphoprotein 1 (Spp1), encoding OPN, was markedly high in the CD153+ cells, whereas its expression was minimal in CD153−PD-1+ and PD-1− cells (Figure 5A). CD153− cells also showed significantly increased expression of cyclin-dependent kinase inhibitor 1A (Cdkn1a, also known as Cip1) and Cdkn2b (also known as Ink4b), which are typical cellular senescence biomarker genes (32), compared with CD153−PD-1+ and PD-1− cells. Both CD153− and CD153−PD-1+ cells showed downregulation of Satb1, eukaryotic translation elongation factor 1α (Eef1a1), and dual-specificity phosphatase 10 (Dusp10) expression, also suggestive of cellular senescence (33, 34). Further, the vast majority of CD153−PD-1+ cells expressed SA–β-gal (Figure 5B) and showed the highest expression of γ-H2AX among the subpopulations (Figure 5C). In 18-week-old HFD-fed mice, CD153−PD-1+CD4+ T cells predominantly accumulated in VAT, although they also appeared in much lower numbers in spleen, liver, and subcutaneous adipose tissue (SAT) and negligibly in blood (Figure 5D). In VAT, CD153− T cells were confirmed to be localized in CLSs (Figure 5E). These results indicated that CD153+PD-1+ CD44hiCD4+ T cells with features of cell senescence were specifically increased and accumulated in VAT of HFD-fed mice.
CD153+PD-1−CD4+ T cells are the main source of OPN in VAT of HFD-fed mice. CD153+PD-1−CD4+ T cells in VAT barely expressed FoxP3, GATA3, or RORγt, but markedly expressed T-bet (Figure 6A), which suggests that these T cells may belong to a Th1 cell lineage. Consistent with this finding, these cells secreted IFN-γ upon TCR stimulation, although the level was significantly lower than that detected in the PD-1+CD4+ T cell population (Figure 6B). However, only CD153+PD-1−CD4+ T cells secreted small yet significant amounts of OPN, even in the absence of TCR stimulation, and the secretion was markedly enhanced through TCR stimulation (Figure 6B). The PD-1+CD4+ T cell population only minimally produced OPN, even after TCR stimulation (Figure 6B), suggesting that the potent OPN production by CD153+PD-1−CD4+ T cells in VAT may not be a mere reflection of the nature of the Th1 cells. To further validate whether these T cells can indeed produce OPN in situ in the VAT of HFD-fed obese mice, we used 18-week-old EGFP-Spp1 knockin (KI) reporter mice fed an HFD on the same schedule. Among CD4+ T cells in VAT, GFP (namely, the activity of Spp1 gene transcription in situ) was expressed almost exclusively in the PD-1+ cell population (Figure 6C, left). A detailed analysis revealed that GFP expression was largely confined to the PD-1hi population, in which the GFP expression level was well correlated with that of CD153; PD-1lo and PD-1null cells expressed minimal and negligible CD153 and GFP, respectively (Figure 6C, right, and Figure 6D). Expression of GFP was undetectable in CD8+ T cells and low in B cells in VAT (Figure 6E). Although macrophages have long been regarded as a source of OPN, expression of GFP was modest in VAT (Figure 6E). Thus, these results suggest that Spp1 gene activation is strongly induced in a proportion of CD153+PD-1+CD4+ T cells in VAT in an unsynchronized manner over a given time period, whereas the Spp1 gene may be activated constitutively to a much lesser extent in VAT macrophages. The PD-1 receptor plays an important role in resolving inflammation by negatively regulating T cell activation upon interaction with the ligands (35–38). We found that adipocytes, macrophages, and B cells, the major antigen-presenting cells in VAT (11, 16, 39), strongly expressed programmed cell death ligand 1 (PD-L1) in HFD-fed mice (Figure 7A). However, although stimulation with solid-phase PD-L–Ig fusion protein significantly suppressed IFN-γ production by CD153+PD-1+CD4+ T cells upon TCR stimulation in vitro, it did not at all affect OPN production by the same cell population (Figure 7B). These results suggest that CD153+PD-1+CD4+ T cells are the main source of OPN in VAT of HFD-fed obese mice, in which these T cells are capable of secreting abundant OPN in a manner resistant to negative signaling via PD-1.

CD153+PD-1−CD4+ T cells induce VAT inflammation and insulin resistance upon cell transfer. To directly examine the possible pathogenic role of CD153+PD-1−CD4+ T cells in VAT inflammation, we performed adoptive transfer of 3 distinct subsets of CD4+ T cells into the VAT of ND-fed lean mice. Since very limited numbers of these cells were obtained after sorting from the VAT, we used splenic CD4+ T cell counterparts populations from 18-week-old HFD-fed obese mice. We confirmed that isolated PD-1-, CD153+PD-1-, and CD153+PD-1+CD4+ T cells from spleens exhibited features that...
were essentially comparable to those from VAT (Supplemental Figure 2, A–C). Our analysis of splenic cells from HFD-fed EGFP- Spp1-KI reporter mice demonstrated that a substantial portion of CD153−PD-1−CD4+ T cells expressed GFP signal at a much higher level (about 2 logs) than did macrophages (Supplemental Figure 2, D–F). Sorted CD4+ T cell subpopulations were mixed with a solution containing a thermoreversible gelation polymer (40) to avoid mechanical cell leakage and transferred directly into the VAT of ND-fed lean mice at 1 × 10^5 cells per mouse under echographic guidance, 3 times per week for 2 weeks, starting at 18 weeks of age. Recipients injected with vehicle alone served as a control group. Immunostaining and flow cytometric analysis confirmed the successful engraftment of injected cells in VAT, without significant leakage around the VAT or other organs, including the liver (Supplemental Figure 3, A–D). Seven days after the last cell transfer, VAT from the recipient mice was analyzed for the expression of inflammation-related and adipocyte-related gene transcripts. VAT from the recipients of CD153−PD-1−CD4+ T cells showed significantly higher expression of Spp1, Ifng, Tnfa, and Il6 and lower expression of Adipoq and Pparg than did the recipients of PD-1+ or CD153−PD-1−CD4+ T cells (Figure 8A). Also, only the recipients of CD153−PD-1−CD4+ T cells exhibited a significant increase in serum OPN and IgG levels compared with control mice (Figure 8B). Furthermore, the recipients of CD153−PD-1−CD4+ T cells and, to a lesser extent, those of CD153−PD-1−CD4+ T cells showed an increase in the proportion of CD11c+CD206+ macrophages relative to CD11c−CD206− macrophages, although CD11c+CD206− macrophages predominated in the recipients of PD-1−CD4+ T cells, similar to what was observed in the control group (Figure 8C). The transfer of any CD4+ T cell subpopulation had no effect on BW, total VAT weight, or food intake (Supplemental Figure 4). However, only the recipients of CD153−PD-1−CD4+ T cells showed significant aggravation of glucose tolerance and insulin sensitivity compared with control mice (Figures 8D). These results suggest that CD153−PD-1−CD4+ T cells are capable of inducing VAT inflammation and metabolic disorder in VAT.

**OPN secreted by CD153−PD-1−CD4+ T cells plays a crucial role in VAT inflammation.** OPN is reported to affect the function of a number of cell types involved in inflammation (41–45). We confirmed that recombinant OPN enhances IFN-γ secretion by both CD8+ and CD4+ T cells and IL-17 secretion by CD4+ T cells via TCR stimulation in culture, while it suppresses IL-10 secretion by CD4+ T cells via TCR stimulation and LPS-stimulated B cells (Supplemental Figure 5, A–C). We then isolated PD-1−, CD153−PD-1+, CD153+PD-1+, and CD4+ T cells from VAT of 18-week-old HFD-fed obese mice and cultured each fraction of cells with B cells from normal B6 mice in the presence of anti-CD3/CD28 mAb. Among the 3 fractions, only TCR-stimulated CD153+PD-1+CD4+ T cells significantly enhanced IgG secretion by B cells, and this secretion was completely abolished in the presence of anti-OPN Ab (Supplemental Figure 5D). An in vitro culture system has inherent difficulties in accurately reproducing true in vivo conditions, yet it shows that CD153−PD-1−CD4+ T cells are capable of inducing VAT inflammation and metabolic disorder in VAT.
chambers and treated with conditioned medium of each type of CD4+ T cells. Comparative analyses for the macrophage migration assay demonstrated that supernatants of CD153+PD-1+CD4+ T cells significantly enhanced macrophage migration, which was inhibited in the presence of anti-OPN Ab (Supplemental Figure 5E). To further confirm the inflammatory role of OPN secreted by CD153+PD-1+CD4+ T cells in VAT, we adoptively transferred CD153+PD-1+CD4+ T cells isolated from the spleens of 18-week-old HFD-fed Spp1−/− and WT B6 mice into VAT of ND-fed lean B6 mice. Although VAT of the recipients of WT CD153+PD-1+CD4+ T cells showed significantly increased expression of the inflammatory genes Spp1, Ifng, Tnfa, and Il6 compared with expression levels in the control VAT, as before, the same CD4+ T cell fraction from Spp1−/− mice hardly induced these effects (Figure 9A). Also, unlike WT CD153+PD-1+CD4+ T cells, the transfer of Spp1−/− CD153+PD-1+CD4+ T cells did not affect the relative proportions of CD11cloCD206hi and CD11chiCD206lo macrophages in VAT (Figure 9B) and failed to induce an increase in OPN and IgG levels in serum (Figure 9C). Further, the recipients of Spp1−/− CD153+PD-1+CD4+ T cells showed no significant aggravation of glucose tolerance or insulin sensitivity, unlike the recipients of WT counterpart cells (Figure 9D). These results suggest that OPN secreted from CD153+PD-1+CD44hiCD4+ T cells plays a crucial role in VAT inflammation and glucose metabolism.

Robust generation of CD153+PD-1+CD4+ T cells in VAT and VAT inflammation in HFD-fed obese mice partly depend on B cells. Finally, we addressed the cellular mechanisms for the accumulation and activation of VAT CD153+PD-1+CD4+ T cells from 18-week-old HFD-fed obese mice. Because obese VAT contained a significant proportion of B cells expressing GL7, a marker of germinal center (GC) B cells (Supplemental Figure 6), we investigated the possible involvement of B cells in the generation of CD153+PD-1+CD4+ T cells in VAT under HFD conditions, using μMT mice that were deficient in mature B cells. HFD-fed μMT mice showed significantly decreased proportions of CD44hiCD62LloCD4+ T cells (Figure 10A), and the effect was associated with diminished numbers of
PD-1+CD4+ T cells and the CD153+ cell fraction in the PD-1+CD4+ T cells in VAT (Figure 10B). In agreement with these findings and consistent with those of a previous report (16), HFD-fed μMT mice showed significantly milder glucose intolerance and insulin resistance than did age-matched HFD-fed WT mice (Figure 11A), although the degrees of weight gain and fat deposition were unchanged (Figure 11B). HFD-fed μMT mice also had assuredly reduced expression of Spp1, Ifng, and Tnfa (Figure 11C) as well as...
a compromised increase in CD11c<sup>hi</sup>CD206<sup>lo</sup> macrophages in VAT compared with HFD-fed WT mice (Figure 11D) and lower levels of OPN in plasma (Figure 11E). These results suggest that B cells play a crucial role in the robust increase of CD153<sup>+</sup>PD-1<sup>+</sup>CD44<sup>hi</sup>CD4<sup>+</sup> T cells in VAT and, accordingly, in VAT inflammation and metabolic disorder under HFD conditions.

Discussion

Various types of immune cells have been identified in VAT, and visceral obesity influences the proliferation and function of these immune cells (7–10). In the current study, we found that a unique subset of CD44<sup>hi</sup>CD4<sup>+</sup> T cells expressing PD-1 and CD153 specifically emerged in the VAT of mice fed an HFD. CD153<sup>+</sup>PD-1<sup>+</sup>CD44<sup>hi</sup>CD4<sup>+</sup> T cells expressed T-bet with negligible GATA3, RORγ<sub>γ</sub>, and FoxP3 expression, suggesting a relationship with the T<sub>h1</sub> cell lineage. Functionally, however, CD153<sup>+</sup>PD-1<sup>+</sup>CD44<sup>hi</sup>CD4<sup>+</sup> T cells preferentially produced large amounts of OPN upon TCR stimulation, while the production of IFN-γ and IL-2 was compromised compared with that detected in PD-1<sup>+</sup>CD4<sup>+</sup> T cells. The vast majority of these T cells also showed a markedly increased expression of SA–β-gal, γ-H2AX, and Cdkn1a/Cdkn2b, with reduced Sath1 expression, suggestive of cellular senescence. Spp1 activation in T<sub>h1</sub> cells is regulated by T-bet, and deregulated OPN production induces excessive T<sub>h1</sub> polarization (46). In addition, T-bet-deficient mice show suppressed proinflammatory immune cell infiltration in VAT and improved glucose homeostasis compared with WT mice (47). It remains to be seen whether the skewed production of large amounts of OPN is due to excessive Th1 polarization or reflects a feature of cellular senescence, namely SA–secretory phenotypes (31). In any case, these T cells were remarkably increased in the VAT of HFD-fed mice, although essentially similar T cells were also detected in much smaller numbers in spleen.

We demonstrated that adoptive transfer of CD153<sup>+</sup>PD-1<sup>+</sup>CD44<sup>hi</sup>CD4<sup>+</sup> T cells, but not other CD4<sup>+</sup> T cells, from HFD-fed spleens into VAT of ND-fed mice recapitulates the features of VAT inflammation, including a striking increase in CD11c<sup>hi</sup>CD206<sup>lo</sup> macrophages and expression of proinflammatory cytokine genes such as Spp1, Ifng, Tnfα, and Il6 in VAT. Importantly, only the recipients of CD153<sup>+</sup>PD-1<sup>+</sup>CD44<sup>hi</sup>CD4<sup>+</sup> T cells developed modest but statistically assured glucose intolerance and increased insulin resistance, strongly suggesting that these unique CD4<sup>+</sup> T cells play a substantial role in initiating VAT inflammation and metabolic disorder in HFD-fed mice.

OPN has been reported to be increased in the circulating blood of obese diabetic and insulin-resistant patients (42, 48) and to play a causative role in VAT inflammation and insulin resistance (19, 49). Using EGFP-Spp1–KI reporter mice, we demonstrated that a significant proportion of CD153<sup>+</sup>PD-1<sup>+</sup>CD44<sup>hi</sup>CD4<sup>+</sup> T cells in situ in VAT of HFD-fed mice showed a remarkable activation of Spp1 in situ that was far greater than that found in macrophages. Adoptive transfer of CD153<sup>+</sup>PD-1<sup>+</sup>CD44<sup>hi</sup>CD4<sup>+</sup> T cells from HFD-fed Spp1<sup>−/−</sup> mice failed to induce VAT inflammation or insulin resistance, indicating a crucial role of the OPN produced by these T cells in VAT.

Nishimura et al. focused on the regulatory B cell (Breg) subset and reported that the proportion and function of Bregs were reduced in HFD-fed VAT, suggesting that Breg dysfunction contributes to the progression of VAT inflammation in obesity (50). OPN activates B cells to produce Igs, while it suppresses IL-10 production by B cells (51). The current study suggests that OPN secreted by CD153<sup>+</sup>PD-1<sup>+</sup>CD44<sup>hi</sup>CD4<sup>+</sup> T cells contributes to the Breg dysfunction we observed in HFD-fed VAT.

In our study, the HFD-fed μMT mice had markedly decreased numbers of CD153<sup>+</sup>PD-1<sup>+</sup>CD44<sup>hi</sup>CD4<sup>+</sup> T cells in VAT, suggesting that B cells were required for their optimal increase. We previously reported that CD153<sup>+</sup>PD-1<sup>+</sup>CD44<sup>hi</sup>CD4<sup>+</sup> T cells in splenic GC cells of lupus-prone mice are activated to produce OPN in response to GC B cells in a TCR/MHC class II–dependent manner (31), and we currently report that obese VAT contains GL-7<sup>+</sup> GC-like B cells. Thus, an intriguing possibility may be that B cells function as antigen-presenting cells to drive the generation of CD153<sup>+</sup>PD-1<sup>+</sup>CD44<sup>hi</sup>CD4<sup>+</sup> T cells in VAT, although the nature of the antigens remains to be elucidated. The B cells and macrophages in VAT were found to strongly express PD-L1 (35–38), a ligand for the negative PD-1 receptor. Notably, however, our study showed that TCR-induced OPN production by CD153<sup>+</sup>PD-1<sup>+</sup>CD44<sup>hi</sup>CD4<sup>+</sup> T cells was hardly affected by PD-1 signaling, suggesting that the activation of these T cells in VAT is not under checkpoint control and allows the chronic progression of VAT inflammation.

Although our study focused on a unique CD4<sup>+</sup> T cell subpopulation in VAT of HFD-fed obese mice, we found that the numbers

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**Figure 7. Adipose CD153<sup>+</sup>PD-1<sup>+</sup>CD4<sup>+</sup> T cells secrete a large amount of OPN independently of the PD-1/PD-L1 inhibitory pathway.** (A) Representative flow cytometric analysis of PD-L1 on adipocytes, macrophages, and B cells obtained from VAT of WT mice fed an HFD (n = 5 mice per group). (B) CD153<sup>+</sup>PD-1<sup>+</sup>CD4<sup>+</sup> T cells were cultured in the presence of anti-CD3/28 Abs along with PD-L1–Ig or control reagents for 3 days. IFN-γ and OPN concentrations in the culture supernatants were assessed by ELISA (n = 5 mice per group). Flow cytometric plots are representative of at least 3 independent experiments. Data represent the mean ± SEM. **P < 0.0001, by 2-tailed Student’s t test.
of CD8+ cells were also significantly increased in VAT of ND-fed mice, with a marked B cell–dependent predominance of CD44hi CD62Llo cells. However, these CD8+ T cells expressed very little CD153, nor did they show activation of the Spp1 gene. Thus, we hypothesize that CD8+ T cells show different features and behave in different ways from CD4+ T cells in HFD-fed VAT. Their exact features remain to be investigated and will be the subject of future research by our group.

It is noteworthy that CD153+PD-1+CD4+ T cells in VAT of HFD-fed mice show features indistinguishable from those of CD153+ SA–T cells, which gradually increase systemically with age (31). The age-dependent increase in CD153+ SA–T cells may partly underlie the immune aging, including a reduction in acquired immunity and an increase in the inflammatory trait and autoimmunity risk (26, 52). Obesity is also associated with diminished resistance against infection (8), chronic low-grade inflammation (53, 54), and a greater susceptibility to autoimmunity (55).

It has been suggested that the increase in CD153+ SA–T cells in chronological aging and systemic autoimmunity is attributable to a robust, homeostatic T cell proliferation (31), but the precise mechanism underlying the accumulation of these T cells in VAT of HFD-fed mice remains to be investigated. Nonetheless, it is an intriguing possibility that the predisposition often associated with obesity may partly be a systemic manifestation of the premature increase in CD153+ SA–T cells in VAT, since adipose tissues can constitute up to 50% to 60% of total BW in severe obesity (7).

In conclusion, we have demonstrated that a unique subpopulation of CD4+ T cells expressing PD-1 and CD153 plays an important role in VAT inflammation and insulin resistance under HFD conditions through OPN production. The current results under-

**Figure 8. Adoptive transfer of CD153+PD-1+CD4+ T cells induces VAT inflammation and insulin resistance in lean mice on an ND.** (A–D) PD-1−, CD153 PD-1−, and CD153+PD-1+CD4+ T cells were isolated from the spleens of WT mice fed an HFD. Next, 1 × 10^6 cells were transferred directly into VAT of individual recipient ND-fed lean mice 3 times per week for 2 weeks, starting at 18 weeks of age. ND-fed lean mice receiving vehicle (thermoreversible gelation polymer) were used as the control group. (A) Real-time PCR analyses of Spp1, Ifng, Tnfa, Il6, and Adipoq, Parg expression levels in VAT of recipient mice (n = 5 mice per group). (B) Analyses of serum OPN and IgG concentrations in recipient mice (n = 5 mice per group). (C) Analyses of CD11c+CD206− macrophage and CD11c+CD206+ macrophage ratios in the VAT of recipient mice (n = 5 mice per group). (D) Effects of adoptive transfer of control reagents or PD-1−, CD153 PD-1−, or CD153+PD-1+CD4+ T cells on the oral glucose tolerance test (OGTT) (C) and insulin tolerance test (ITT) (D) (n = 5 mice per group).

*P < 0.05, for CD153+PD-1+CD4+ T cells versus vehicle alone. Data represent the mean ± SEM. *P < 0.05, **P < 0.001, and ***P < 0.0001, by ANOVA followed by post hoc Bonferroni tests.
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score a possible link between visceral obesity and immune aging and may provide a cellular target for controlling immunometabolic anomalies in visceral obesity.

Methods
Animal work. C57BL/6 (B6) mice were purchased from Japan SLC Inc., and μMT, Spp1-KO, and EGFP-Spp1-KI reporter mice were described before (31). All mice were housed under a 12-hour light/12-hour dark cycle and allowed free access to food. Mice were fed with either a ND (CE-2, 6 kcal % fat; CLEA Japan Inc.) or an HFD (D12492, 60 kcal % fat; Research Diets Inc.). We used littermate controls in the experiments involving Spp1-KO and EGFP-Spp1-KI reporter mice on a B6 background. B6 mice were used as a control for μMT mice (which were purchased from The Jackson Laboratory), according to The Jackson Laboratory’s recommendation.

Isolation of the stromal vascular fraction and flow cytometry. We isolated stromal vascular cells using previously described methods, with some modifications. Mice were sacrificed under general anesthesia after systemic heparinization. VAT was removed and ground into small pieces. Samples were incubated for 40 minutes in collagenase II/DNase I solution (1 mg/ml collagenase II and 50 μg/ml in HBSS solution), with gentle stirring. Digested tissue was then centrifuged at 1,000 g for 10 minutes. The resulting pellets were washed twice with cold PBS and filtered through a 70-mm mesh. Red blood cells were lysed with erythrocyte-lysing buffer (eBioscience) for 5 minutes and resuspended in RPMI-1640 supplemented with 10% FBS. Single-cell suspensions of splenocytes and the adipose stromal vascular fraction (SVF) were blocked with CD16/32 mAb (2.4G2; BD Biosciences) at 4°C for 5 minutes. Cells were stained with a mixture of Abs at 4°C for 20 minutes. Flow cytometric analysis and cell sorting were performed on a FACSaria III instrument (BD Biosciences) and analyzed using FlowJo software. The purity of the sorted cells was consistently more than 95% by post-sorting analysis. The representative post-sorting analyses are shown in Supplemental Figure 7.
The Abs used were specific to CD3 (14A-2; BioLegend); CD4 (GK1.5; BioLegend); CD8a (53-6.7 and OKT8; eBioscience); CD44 (IM7; BioLegend); CD62L (MEL-14; eBioscience); CD11b (M1/70; eBioscience); F4/80 (BM8; BioLegend); CD11c (N418; eBioscience); CD206 (MR5D3; Biolegend); PD-1 (RMP1-30 and 29F.1A12; BioLegend); CD45 (30F11.1; eBioscience); CD19 (1D3; eBioscience); PD-L1 (10F.9G2; BioLegend); CD153 (RM153; BioLegend); GL7 (GL7; BioLegend); and 7-AAD viability staining solution (BioLegend). We analyzed intracellular cytokine production with a Cytofix/Cytoperm Fixation/Permeabilization Solution Kit (BD Pharmingen) and a transcriptional factor using a True-Nuclear Transcription Factor Buffer Set (BioLegend). The Abs used were specific to FoxP3 (FJK-16s; eBioscience); T-bet (4B10; BioLegend); GATA3 (16E10A23; BioLegend); RORγt (B2D; eBioscience); and γ-H2AX (N1-431; BD Pharmingen).

**IHC.** Whole-mount VAT was stained and visualized. Mice were sacrificed by cervical dislocation, after which VAT was removed using sterile technique and minced into small pieces (~2–3 mm) using a scalpel. Tissue pieces were washed, fixed in cellFIX (catalog 340181; BD) for 60 minutes, and permeabilized with 0.1% Triton X-100 for 10 minutes. The specimens were then blocked with 5% BSA and incubated with primary Abs specific to CD3 (145-2C11; BD); PD-1 (J43; Bio X Cell); F4/80 (BM-8; eBioscience); and CD153 (RM153; eBioscience) for 1 hour and then with Alexa Fluor 488- and 647-conjugated secondary Abs (Molecular Probes, Thermo Fisher Scientific) for 1 hour. The vasculature was stained with *Griffonia simplicifolia* B4 isoelectin conjugated with Alexa Fluor 488 (FL-1201; Vector Laboratories) for 1 hour. Tissue samples were excited using 4 wavelengths (405, 488, 568, and 800 nm) and the emission collected through the appropriate narrow band-pass filters on a confocal microscope (LSM 510 META; Carl Zeiss). The images were acquired and processed using LSM 510 software (Carl Zeiss).

**Quantitative real-time PCR.** Total RNA samples from sorted cells and VAT were prepared using an RNeasy Mini Kit (QIAGEN) or TRIzol reagent (Invitrogen, Thermo Fisher Scientific) according to the manufacturers’ instructions. A First-Strand cDNA Synthesis Kit (Invitrogen, Thermo Fisher Scientific) was used for cDNA synthesis. Quantitative real-time PCR was performed using the ABI PRISM 7700 Sequence Detection System (Applied Biosystems) or the ViiA 7 Real-Time PCR System (Thermo Fisher Scientific). The *Gapdh* gene was used as an endogenous control to normalize for differences in the amount of total RNA in each sample. All values were expressed as a fold increase or decrease relative to the expression of *Gapdh*. The primer sequences for genes were as follows: *Gapdh*, 5′-AGGTCGGTGTAACGGATTTG and 3′-TGTAGACCATGTAGTTGAGGTCA; *Spp1*, 5′-CCCGTGAGTGCTGATT and 3′-TTCTTCAGAGGACACAGCATTC; *Ifng*, 5′-ATCTGGAGGAACTGGCAAAA and 3′-TTCAAGACTTCAAGAGTCTGAGG; *Cebpa*, 5′-TGAGAAAAATGAAGGGTGCAG and 3′-CGGGATCTCAGCTTCCTGT; *Satb1*, 5′-ACTGAAACGAGCCGAATC and 3′-CGGAGGATTTCAGAAAGCAA; *Tnfα*, 5′-CCCTCACTCAGATCTTCTGT and 3′-GCTACGACGTGGGCTACAG; *Eef1a1*, 5′-CAACATCGTGTAATCGGACTT and 3′-TGTAGACCATGTAGTTGAGGTCA; *Dusp10*, 5′-CCATCTCCTTTAGACGACGG and 3′-GCTACCACTACCTGGGCTG; *Emr1*, 5′-TTTGCAGTGCAACTCAGGACT and 3′-GATCCCCAGGTGTGATGCA; *Pparg*, 5′-TCCGCGTGACGGATGCTAT and 3′-GAGAGGTC-
B cell culture. VAT B cells (1 × 10^5) were purified by positive selection using anti-CD19 MACS Beads (Miltenyi Biotec) according to the manufacturer’s instructions and were generally more than 95% CD19+B220+. B cells were cultured in DMEM supplemented with 10% FBS plus LPS from *Escherichia coli* (1 μg/ml; Sigma-Aldrich) in the presence or absence of recombinant OPN for 72 hours. We used 3 mice per sample of the cells.

ELISA. The levels of OPN (R&D Systems); IFN-γ (BioLegend); IL-17 (R&D Systems); and total IgG (eBioscience) in supernatants or serum were determined by ELISA according to the manufacturers’ instructions.

Migration of peritoneal macrophages. We isolated 1 × 10^5 PD-1−, CD153−PD-1+, and CD153+PD-1+CD4+ T cells from VAT of HFD-FED WT mice. We used 5–7 mice per sample of the cells. These cells were cultured in complete RPMI with immobilized anti-CD3 mAb (5 μg/ml) and soluble anti-CD28 mAb (2.5 μg/ml). After 120 hours of culture, the culture medium was aspirated and analyzed by a migration assay using Boyden chambers with 8-mm pore inserts (BD). Peritoneal macrophages were cultured in the upper wells, and the conditioned medium was added to the lower wells. We used fresh DMEM supplemented with 5% FBS as a control. To inhibit OPN activity, an anti-OPN Ab (2 μg/ml; Sigma-Aldrich) or control IgG was added to the conditioned medium.

Adoptive transfer of T cells into recipient mice. PD-1−, CD153–PD-1+, and CD153+PD-1+CD4+ T cells were sorted from the spleens of HFD-fed WT mice (Figure 5), and CD153+PD-1+CD4+ T cells were sorted from the spleens of WT or *Spp1*−/− mice fed an HFD (Figure 6). Sorted CD4+ T cells were mixed with a thermoreversible gelation polymer CACAGAGCTGATT; *Adipoq*, 5′-TGTTCCTCTTAATCCTGCCCA and 3′-CCAACCTGGCAAAAGTCCT; Tgfb1, 5′-CTCCCGTG - GCTGAGCTCTAC and 3′-GCCCTTTGAGGATGCTG; Cdkn1a, 5′-AACATCTCAGGGCCGAAA-3′ and 5′-TGCGCTTGGAGTGA - TAGAAA-3′; and Cdkn2b, 5′-AATAACTTCCTACGCATTTTCTGC-3′ and 5′-CCCTTGGCTTCAAGGTGAG-3′.

Glucose and insulin tolerance tests. We performed glucose tolerance (oral administration of 1.5 g/kg of glucose, after 16 hours of fasting) and insulin tolerance (administration of 0.75 U/kg of insulin i.p. after 4 hours of fasting) tests to assess glucose intolerance and insulin resistance.

**B cell helper assay.** B cells (3 × 10^5) from VAT and spleen were purified by positive selection using anti-CD19 MACS Beads (Miltenyi Biotec) according to the manufacturer’s instructions; the B cells were generally more than 95% CD19+B220+. For coculture experiments, PD-1−CD4+, CD153−PD-1+, and CD153+PD-1+CD4+ T cells were sorted from VAT or spleens of WT mice fed an HFD. These were stimulated with 5 μg/ml plate-bound anti-CD3 with anti-OPN Ab (2 μg/ml; Sigma-Aldrich) or control IgG and mixed with B cells at a 1:3 T cell/B cell ratio in U-bottom 96-well plates. We used 5–7 mice per sample of the indicated cells (Supplemental Figure 5D). Ten days later, the total IgG concentration of the supernatants was determined by ELISA (eBioscience).

**T cell culture.** CD4+ T cells or CD8+ T cells (1 × 10^5) were stimulated with immobilized anti-CD3 mAb (5 μg/ml) with soluble anti-CD28 mAb (2.5 μg/ml) in the presence or absence of 1 μg/ml mouse recombinant OPN for 72 hours. We used 5–7 mice per sample of the cells. PD-L1-Ig or control IgG (R&D Systems) was additionally included in the culture.
(Mebiol Gel; Cosmo Bio). We transferred $1 \times 10^5$ cells into VAT of recipient mice fed an ND directly under echographic guidance, 3 times per week for 2 weeks, starting at the indicated age (Figure 8 and 9). The vehicle (thermoresversible gelation polymer) was similarly injected into the control mice.

**Cellular senescence assay.** We isolated the SVF from VAT and splenocytes of HFD-fed WT mice. SA-β-gal assays were performed using a Cellular Senescence Live Cell Analysis Kit (Enzo Life Sciences).

**Statistics.** All values are presented as the mean ± SEM. The statistical significance of differences between 2 groups was determined using a 2-tailed Student’s t test. Differences among multiple groups were compared using ANOVA followed by Bonferroni’s post-hoc tests. A P value of less than 0.05 was considered significant.

**Study approval.** This study conformed to NIH guidelines (Guide for the Care and Use of Laboratory Animals. National Academies Press. 1996.) and was approved by the IACUC of the Keio University School of Medicine.

**Author contributions**
K. Shirakawa conducted most of the experiments. KY obtained the preliminary results. K. Shimnura, JE, MK, YK, TY, AA, NY, and M. Sekai and SI helped with the experimental work. HI, IM, YH, and KF supervised the study. M. Sano and K. Shimnura designed the experiments. NM and M. Sano wrote the manuscript.

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