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Foods high in fermentable oligosaccharides, disaccharides, monosaccharides, and polyols (FODMAPs) exacerbate symptoms of irritable bowel syndrome (IBS); however, their mechanism of action is unknown. We hypothesized that a high-FODMAP (HFM) diet increases visceral nociception by inducing dysbiosis and that the FODMAP-altered gut microbial community leads to intestinal pathology. We fed rats an HFM and showed that HFM increases rat fecal Gram-negative bacteria, elevates lipopolysaccharides (LPS), and induces intestinal pathology, as indicated by inflammation, barrier dysfunction, and visceral hypersensitivity (VH). These manifestations were prevented by antibiotics and reversed by low-FODMAP (LFM) diet. Additionally, intracolonic administration of LPS or fecal supernatant (FS) from HFM-fed rats caused intestinal barrier dysfunction and VH, which were blocked by the LPS antagonist LPS-RS or by TLR4 knockdown. Fecal LPS was higher in IBS patients than in healthy subjects (HS), and IBS patients on a 4-week LFM diet had improved IBS symptoms and reduced fecal LPS levels. Intracolonic administration of FS from IBS patients, but not FS from HS or LFM-treated IBS patients, induced VH in rats, which was ameliorated by LPS-RS. Our findings indicate that HFM-associated gut dysbiosis and elevated fecal LPS levels induce intestinal pathology, thereby modulating visceral nociception and IBS symptomatology, and might provide an explanation for the success of LFM diet in IBS patients.

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FODMAP diet modulates visceral nociception by lipopolysaccharide-mediated intestinal inflammation and barrier dysfunction

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Foods high in fermentable oligosaccharides, disaccharides, monosaccharides, and polyols (FODMAPs) exacerbate symptoms of irritable bowel syndrome (IBS); however, their mechanism of action is unknown. We hypothesized that a high-FODMAP (HFM) diet increases visceral nociception by inducing dysbiosis and that the FODMAP-altered gut microbial community leads to intestinal pathology. We fed rats an HFM and showed that HFM increases rat fecal Gram-negative bacteria, elevates lipopolysaccharides (LPS), and induces intestinal pathology, as indicated by inflammation, barrier dysfunction, and visceral hypersensitivity (VH). These manifestations were prevented by antibiotics and reversed by low-FODMAP (LFM) diet. Additionally, intracolonic administration of LPS or fecal supernatant (FS) from HFM-fed rats caused intestinal barrier dysfunction and VH, which were blocked by the LPS antagonist LPS-RS or by TLR4 knockdown. Fecal LPS was higher in IBS patients than in healthy subjects (HS), and IBS patients on a 4-week LFM diet had improved IBS symptoms and reduced fecal LPS levels. Intracolonic administration of FS from IBS patients, but not FS from HS or LFM-treated IBS patients, induced VH in rats, which was ameliorated by LPS-RS. Our findings indicate that HFM-associated gut dysbiosis and elevated fecal LPS levels induce intestinal pathology, thereby modulating visceral nociception and IBS symptomatology, and might provide an explanation for the success of LFM diet in IBS patients.
Results

HFM increases mucosal inflammation, gut permeability, and visceral hyperalgesia in the colon. To study the impact of HFM on mucosal inflammatory tone, gene expression of inflammatory cytokines was measured in colonic tissues. IL-1β, IL-6, IL-17, TNF-α, and IFN-γ mRNA levels increased significantly in HFM rats, compared with regular chow–fed (RC-fed) rats ($n=6$, $P<0.05$, Student’s $t$ test) (Figure 1A).

Intestinal barrier function was evaluated by measuring the transepithelial electrical resistance (TEER) of ex vivo colonic tissues. TEER reflects paracellular resistance imparted by tight junctions and the lateral paracellular space and is a sensitive measure of barrier integrity. Intestinal resistance was reduced in HFM rats compared with RC rats ($n=6$, $P<0.05$) (Figure 1B). A reduced TEER was accompanied by reduced expression of the epithelial tight junction proteins zonula occludens–1 (ZO-1) and occludin (OCLN) by 54.0% ± 13.4% and 35.3% ± 11.8%, respectively ($n=6$) (Figure 1, C and D). In vivo assessment of gut permeability revealed a significant increase in serum FITC–dextran in HFM rats ($n=6$, $P<0.05$), indicating impairment of intestinal barrier function (Figure 1E).

Serum LPS in rats fed RC or HFM was measured. The LPS concentration was significantly increased in HFM rats, compared with RC rats (0.84 ± 0.10 vs. 0.40 ± 0.08 endotoxin units [EU]/ml, $n=6$, $P<0.05$) (Figure 1F). These data indicate that impaired

ing lamina propria and deeper intestinal layers. Evidence shows that increased LPS levels compromise intestinal tight junction permeability (15), thus allowing toxic substances such as endotoxins and microorganisms to cross the intestinal wall.

We hypothesize that FODMAPs cause endotoxemia and evoke intestinal inflammation, which in turn modulates visceral nociception. To test this hypothesis, we conducted experiments in rats to determine if a diet with high levels of FODMAPs can cause dysbiosis and lead to the production of endotoxins that cause gut inflammation and induce visceral hypersensitivity. In separate studies, we examined the effects of fecal supernatant obtained from rats fed a high-FODMAP (HFM) diet on visceral sensitivity in naive rats and the permeability of cultured human colonoids. We performed intervention studies in 2 chronic stress rat models to determine if FODMAP restriction prevents low-grade mucosal inflammation and improves gut barrier function.

To further test our hypothesis, we examined the effects of fecal supernatant obtained from IBS-D patients before and after 4 weeks of LFM diet treatment on visceral sensitivity in naive rats. The results were compared to HS. We conclude that increases in fecal LPS after ingestion of an HFM diet and in IBS-D patients mediate intestinal barrier dysfunction and visceral hypersensitivity due to dysbiosis. These abnormalities can be prevented by an LFM diet.
mucosal permeability was accompanied by increased serum LPS.  

Histological analysis revealed increased numbers of mononuclear cells, neutrophils, eosinophils, and mast cells in the lamina propria of the colon from HFM compared with RC rats, suggesting low-grade mucosal inflammation after HFM exposure (Supplemental Table 1; supplemental material available online with this article; https://doi.org/10.1172/JCI92390DS1).

Lastly, the effect of HFM on visceral motor response (VMR) to graded colorectal distension (CRD) was examined. Rats showed pressure-dependent increases in VMR to CRD (Figure 2, A and B). The VMR to 20, 40, 60, and 80 mmHg CRD significantly increased 2.5-, 2.8-, 2.5-, and 2.4-fold after 2-week HFM (P < 0.05, n = 6 per group), suggesting the development of visceral hypersensitivity.

Rifaximin prevents HFM-induced mucosal inflammation, impaired gut permeability, and visceral hyperalgesia. To determine if gut mucosal changes and development of visceral hypersensitivity are caused by HFM acting directly on the mucosa or indirectly due to bacterial community changes in the gut, we investigated the effects of rifaximin, which has been shown to alter bacterial communities and prevent stress-induced gut inflammation and visceral hyperalgesia in rats (16).

Oral gavage of rifaximin prevented an increase in the numbers of colonic mucosal inflammatory cells (Supplemental Table 1) and elevated IL-1β, IL-6, IL-17, TNF-α, and IFN-γ mRNA levels in HFM rats (P < 0.05) (Figure 1A). Rifaximin also prevented a reduction in ZO-1 and OCLN (Figure 1, C and D) and a decrease in TEER in the colon (P < 0.05) (Figure 1B) and impairment of intestinal barrier functions measured by serum FITC–dextran (Figure 1E). Rifaximin also blocked the increase in serum LPS (Figure 1F).

We next assessed the preventive effect of rifaximin on the development of visceral hyperalgesia in HFM rats. Repeated oral gavage of rifaximin significantly attenuated the increased VMR to CRD induced by HFM at 20, 40, 60, and 80 mmHg (Figure 2, A and B). These data suggest that HFM causes mucosal inflammation and visceral hypersensitivity by inducing dysbiosis, which rifaximin prevents.

HFM diet alters the bacterial community. The Illumina MiSeq sequencing platform was used to determine if HFM altered the luminal microbial communities of the ascending colon. The major phyla identified in the ascending colon of rats fed RC were Actinobacteria, Bacteroidetes, Firmicutes, and Verrucomicrobia (Figure 3A). Firmicutes significantly decreased after HFM (Figure 3B). In contrast, after HFM, the relative abundance of Actinobacteria (Gram-positive) and Verrucomicrobia (Gram-negative) significantly increased (Figure 3B). There was an enrichment in the abundance of Gram-negative bacteria after HFM and reduction of the Gram-positive (Figure 3, C and D). Among Gram-negative bacteria, Verrucomicrobia (Akkermansia muciniphila) was increased by 4.7-fold from 8.2% ± 6.5% to 38.3% ± 5.3% (percentage of total Gram-negative bacteria) (Figure 3E).

Effects of rifaximin on HFM-induced microbiota changes and mucosal inflammation. Rifaximin was shown to alter bacterial communities in the ascending colon. Compared with HFM, the rats given rifaximin had a significantly higher relative abundance of the Firmicutes (Figure 3B). There was also a significantly lower abundance of the Actinobacteria and Verrucomicrobia in the rifaximin group compared with HFM (Figure 3, B and E).

Elevated LPS in fecal samples from HFM rats. To determine whether gut dysbiosis results in an increase in fecal LPS levels, the fecal samples from rats fed RC or HFM were obtained and endotoxin (LPS) levels were measured. Fecal LPS measurements showed an endotoxin level approximately 2-fold higher in HFM compared with RC rats (8.6 vs. 5.1 EU/mg fecal content, n = 6, P < 0.05) (Figure 4A).

Effects of fecal supernatant from HFM rats on intestinal barrier function and visceral sensitivity. To evaluate the effects of fecal material on intestinal barrier function and visceral sensitivity, fecal supernatant (0.3 ml) from HFM or RC rats was administered intracolonically to naïve rats. Colonic mucosa inflammatory tone and permeability were measured, and the VMR to CRD was performed 4–6 hours after supernatant administration.

Intracolonic administration of supernatant from HFM rats increased gene expression of IL-1β, IL-6, and TNF-α in the ascending colon compared with administration of fecal supernatant from RC rats (Student’s t test, n = 6, P < 0.05) (Figure 4B). It also reduced gene expression of the epithelial tight junction proteins ZO-1 and OCLN (Figure 4, C and D). Intracolonic HFM supernatant caused intestinal barrier dysfunction by decreasing TEER in the colonic mucosa, compared with RC rats (29.9 ± 0.85 vs. 36.2 ± 0.9 Ω/cm², n = 6, P < 0.05) (Figure 4E) and 2- to 4-fold increases in VMR to CRD at 20, 40, 60, and 80 mmHg (Figure 4F). A potent LPS antagonist, LPS-RS (100 μg/kg), prevented an increase in mucosal cytokine expression, intestinal barrier dysfunction, and visceral hypersensitivity caused by HFM (Figure 4, C–F). The effects of LPS-RS are
Human colonoids were injected with fecal supernatant containing FITC–dextran, and fluorescence images were obtained at different time points. When exposed to fecal supernatant from RC rats, the colonoids retained 85% of FITC–dextran at 12 hours. LPS reduced FITC–dextran retention to 40% ($n = 8$, $P < 0.05$) (Figure 5). Similarly, fecal supernatant from HFM rats reduced FITC–dextran retention to 43% ($n = 8$, $P < 0.05$). The increase in permeability evoked by fecal supernatant from HFM rats was prevented by coadministration of LPS-RS with fecal supernatant from HFM rats (Figure 5).

LFM prevents restraint stress–induced mucosal inflammation, intestinal barrier impairment, and visceral hyperalgesia. A diet with restricted FODMAP content has been reported to reduce symptoms in IBS patients (3, 5). We used a rodent restraint stress (RS) model to determine if LFM prevents intestinal inflammation and visceral hypersensitivity in RS rats.

RS showed evidence of mucosal inflammation characterized by increased numbers of neutrophils and mononuclear cells (Supplemental Table 2) accompanied by increased gene specific, since intracolonic administration of LPS-RS (1.0 mg/kg, 0.3 ml) failed to block visceral hypersensitivity induced by intracolonic infusion of protease-activated receptor 2 (PAR2) agonist, SLIGKV-NH2 (0.5 mg/kg, intracolonically) (Supplemental Figure 1). Separate studies showed that intracolonic administration of LPS (10 μg/kg, 0.3 ml) impaired colonic permeability (Figure 4G) and induced an increase in VMR to CRD, which was prevented by LPS-RS (100 μg/kg, 0.3 ml) (Figure 4H).

To examine whether TLR4 is involved, rat colon was pretreated with the intracolonic administration of siRNA targeting TLR4 (10 nmol per rat in 0.3 ml) for 3 days, which resulted in a 40% reduction in TLR4 gene expression in the ascending colonic mucosa (Supplemental Figure 2). Pretreatment of TLR4 siRNA prevented visceral hypersensitivity caused by fecal supernatant from HFM rats, indicating the involvement of TLR4 (Figure 4F).

Effects of fecal supernatant from HFM rats on epithelial barrier function of cultured human colonic organoids. We used inside-out human colonoids to test the permeability of the colonic epithelium. Human colonoids were injected with fecal supernatant containing FITC–dextran, and fluorescence images were obtained at different time points. When exposed to fecal supernatant from RC rats, the colonoids retained 85% of FITC–dextran at 12 hours. LPS reduced FITC–dextran retention to 40% ($n = 8$, $P < 0.05$) (Figure 5). Similarly, fecal supernatant from HFM rats reduced FITC–dextran retention to 43% ($n = 8$, $P < 0.05$). The increase in permeability evoked by fecal supernatant from HFM rats was prevented by coadministration of LPS-RS with fecal supernatant from HFM rats (Figure 5).
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0.04 ± 0.01 μg/ml, n = 6, P < 0.05, Student’s t test) (Figure 6C).

The serum LPS level was significantly increased in RS compared with control rats (0.58 ± 0.04 vs. 0.38 ± 0.02 EU/ml, n = 6, P < 0.05), indicating metabolic endotoxemia (Figure 6D). Western blot analysis of the tight junction protein ZO-1 in proximal colon

Figure 4. Effects of fecal supernatant from HFM rats and LPS on gut permeability and visceral sensitivity. (A) Endotoxin level (LPS) in fecal contents was elevated in HFM compared with RC rats (n = 6) *P < 0.05. (B–D) Intracolonic infusion of fecal supernatant from HFM rats caused changes in cytokines and junction proteins, ZO-1 and OCLN. These changes were prevented by LPS antagonist, LPS-RS. (E) HFM fecal supernatant decreased the transepithelial electrical resistance (TEER), indicating an increase in epithelium permeability of colon mucosa in naive rats (n = 6, P < 0.05). This increase was prevented by LPS-RS. (F) Intracolonic infusion of fecal supernatant from HFM rats induced visceral hyperalgesia in naive rats; this was prevented by LPS-RS or siRNA targeting TLR4 (n = 6, P < 0.05). (G) Intracolonic LPS infusion induced increased epithelium permeability of colonic mucosa (i.e., reduced TEER) in naive rats; this was prevented by LPS-RS. (H) Intracolonic infusion of LPS (10 μg/kg) induced visceral hyperalgesia in naive rats; this was prevented by LPS-RS (100 μg/kg). *P < 0.05 versus RC or PBS treated; #P < 0.05 versus HFM-supernatant or LPS treated. AUC, area under the curve; EMG, electromyographic activity; HFM, high-FODMAP diet; LPS, lipopolysaccharide; RC, regular chow; siRNA, small (or short) interfering RNA; TLR4, Toll-like receptor 4.

expression of IL-6 (1.3 ± 0.1-fold), IL-17 (1.4 ± 0.1-fold), TNF-α (1.5 ± 0.2-fold), and IFN-γ (1.5 ± 0.4-fold) (Figure 6A). This was accompanied by a 17% decrease in TEER (n = 6, P < 0.05) (Figure 6B). In vivo assessment of gut permeability revealed a significant increase in serum FITC–dextran in RS rats (0.19 ± 0.03 vs. 0.04 ± 0.01 μg/ml, n = 6, P < 0.05, Student’s t test) (Figure 6C).
tissues showed a significant decrease in protein expression after chronic RS (n = 6, P < 0.05) (Figure 6E), indicating impairment of intestinal barrier function after chronic stress.

Two-week LFM feeding prevented RS-provoked cytokine gene expression (Figure 6A). LFM normalized the RS-induced increase in TEER (35.4 ± 1.5 vs. 41.0 ± 1.2 Ω/cm², n = 6, P < 0.05) (Figure 6B) and the in vivo increased gut permeability to FITC-dextran (n = 6, P < 0.05) (Figure 6C). In addition, LFM prevented an increase in serum LPS (Figure 6D), reduction of ZO-1 protein (Figure 8E), and development of visceral hypersensitivity (Figure 8E). These changes were prevented by LFM (Figure 8A–F).

**Patient demographics and IBS symptoms before and after LFM.** Six IBS-D patients (4 females and 2 males), ages ranging from 43 to 59 years old, were recruited from the University of Michigan GI outpatient clinic and primary care clinics. Six sex- and age-matched HS were recruited to serve as controls. All 6 IBS-D subjects were symptomatic, with symptom scores presented in Table 1. Following 4 weeks of LFM, all 6 patients experienced significant improvement of abdominal pain and bloating (Table 1). There was a trend towards symptomatic improvement with stool frequency, consistency, and urgency, but the differences did not reach statistical significance (Table 1).

**Effects of fecal supernatant from IBS-D patients on visceral sensitivity of naïve rats.** Endotoxin assays were performed on fecal samples from 6 HS and 6 IBS-D patients. These assays revealed a higher level of fecal LPS among IBS-D patients compared with HS (16.2 EU/μg vs. 7.9 EU/μg, P < 0.05). Four-week treatment with LFM diet lowered fecal LPS to a level similar to that of HS (Figure 9A).

To evaluate the effects of fecal material on visceral sensitivity, fecal supernatant (0.3 ml) from each of the 6 IBS-D patients and each of the 6 HS was intracolonically administered to the colon of naive rats and then the VMR to CRD was evaluated 4–6 hours after supernatant administration. Intracolonical injection of IBS fecal supernatant evoked visceral hypersensitivity and caused an increase in VMR to CRD at 20, 40, 60, and 80 mmHg compared with fecal supernatant from HS (Figure 9B and C). In contrast, the VMR to intracolonial administration of fecal supernatant from IBS-D patients following 4-week LFM diet caused an increase in VMR to CRD, which was similar to that observed...
in HS (Figure 9, B and C). Furthermore, coadministration of the LPS antagonist LPS-RS (100 μg/kg, intracolonically) prevented the development of visceral hypersensitivity caused by fecal supernatant from IBS-D, which indicates that elevated fecal LPS is responsible for the induction of visceral hypersensitivity. Thus, similar to our rat data, our human studies suggest that IBS-D patients have elevated fecal LPS levels resulting from gut dysbiosis and this appears to be responsible for the development of visceral hypersensitivity in IBS-D patients.

Discussion

Food is associated with symptom onset or exacerbation in many IBS patients. Ingestion of sugar (lactose or fructose), sorbitol, and oligosaccharides (fructans) alone or in combination may trigger abdominal symptoms (1, 17, 18). However, the mechanisms responsible for symptom generation in IBS remain poorly understood. In general, luminal water retention by osmotically active short-chain carbohydrates and increased gas production due to bacterial fermentation are believed to induce gastrointestinal symptoms in patients with visceral hypersensitivity (1).

Our study shows for the first time to our knowledge that HFM induces mucosal inflammation and impaired gut permeability. These changes are similar to subclinical mucosal inflammation and increased permeability in a subset of IBS patients (19, 20). Furthermore, we also observed a reduction of ZO-1 and OCLN gene expression in the colon of rats receiving an HFM. A similar reduction of ZO-1 and OCLN has been reported in IBS patients (21). It is conceivable that other tight junction proteins such as claudin-1, -3, -4, -5, and -8 that decrease paracellular permeability and claudin-2 that forms charge-selective paracellular pores could also be affected by HFM and this should be evaluated in future studies. In addition, the recognized correlation between impaired gut permeability and visceral hypersensitivity (22) was also observed in HFM rats. A cause-and-effect relationship between mucosal barrier alteration and visceral hypersensitivity (23) has been described in a rodent model in which chemical blockade of enhanced stress-induced paracellular permeability was accompanied by reduced sensitivity to colonic distension (23). It is conceivable that HFM-induced endotoxemia contributes to hyperexcitability of dorsal root ganglia innervating the distal colon.

An increased serum LPS concentration has been reported in patients with IBS-D compared with HS (24). Endotoxins, which are a component of the outer membrane of Gram-negative bacteria, stimulate various inflammatory mediators. It is conceivable that LPS from Gram-negative bacteria gain access to the gut...
Recent studies reported that Akkermansia muciniphila has a deleterious effect on dextran sulfate sodium–induced colitis (31). Furthermore, LPS purified from Akkermansia muciniphila produces a strong inflammatory response characterized by the production of the NF-κB–dependent cytokines IL-10, TNF-α, IL-1β, and IL-6 similar to that elicited by Escherichia coli (32). In human studies, it has been reported that an increased number of Akkermansia muciniphila in IBS patients is closely correlated with episodes of abdominal pain (33, 34).

Intracolonic administration of fecal supernatant from HFM rats impaired colonic permeability, as reflected by reduced TEER, and increased leakage of fluorescent dye in human colonoids. The VMR to CRD was also increased. These abnormalities were prevented by coadministration of the LPS antagonist LPS-RS. Hence, HFM likely induces gut dysbiosis, causing an increase in fecal LPS, which may impair gut barrier function and induce visceral hypersensitivity.

LPS from Gram-negative bacteria bind to the LPS-binding protein coupled to CD14 on the cell surface. LPS–CD14 interacts with TLR4 to form a complex with another accesso-
To determine if HFM and its metabolites induce mucosal changes and visceral hypersensitivity through direct actions on the gut epithelium and its immune system or through gut dysbiosis, we examined the effects of rifaximin, a nonabsorbable, broad-spectrum antibiotic. We have previously shown amelioration of mucosal inflammation and normalization of visceral hypersensitivity following rifaximin treatment in rats subjected to WAS by modulating host gut bacterial communities (16). In the current study, we show that rifaximin prevents HFM-induced mucosal inflammation, impaired gut permeability, and visceral hypersensitivity. It should be noted that under certain experimental conditions, rifaximin may exert its effects independently of its ability to alter the microbial composition. It was reported that rifaximin therapy ameliorated gut inflammation and endotoxemia in mice treated with humanized microbiota (36). This occurred without alteration in the relative microbial composition. However, it is well known that rifaximin causes a reduction
in the total fecal bacteria count (16). Our studies show that in rats fed an HFM, rifaximin therapy reduced endotoxemia accompanied by a decrease in Gram-negative bacteria such as Verrucomicrobia. Hence, our data suggest that HFM causes these changes by inducing dysbiosis, which rifaximin prevents. Although LFM appears to be effective in the treatment of IBS, little is known about the mechanism. Recognizing the lack of suitable IBS animal models, we chose 2 rat models of visceral hyperalgesia, and mucosal inflammation induced by RS or repeated WAS, to determine if LFM feeding alters gut microbiota, prevents subclinical intestinal inflammation, improves gut barrier function, and reduces visceral hyperalgesia. Psychological stressors have been shown to alter the bacterial community. Previous research using 454 pyrosequencing has shown that stress caused by prolonged restraint significantly changes the bacterial community structure, reduces species richness and diversity, and facilitates host susceptibility to a bacterial pathogen (37). Social stress also significantly alters bacterial community composition and decreases species richness and diversity (38). We have shown that chronic stress caused a significant loss of several bacterial family groups and decreased community diversity and evenness (16). These changes in bacterial community composition can contribute to mucosal barrier impairment and inflammation. In the current study, rats subjected to RS showed an increased relative abundance of Gram-negative bacteria, as well as an increase in LPS levels in the colon, accompanied by mucosal inflammation and impaired gut permeability. These abnormalities are similar to those observed in IBS patients (17, 19, 20). In the current study, we show for the first time to our knowledge that 2-week LFM prevents the development of mucosal inflammation, impaired permeability, and visceral hypersensitivity evoked by stress. Interestingly, LFM also prevents the increase in colonic Gram-negative bacteria and LPS. This suggests that LFM acts by modulating the gut microbiota, which seems to be affected by stress. This is not surprising, as HFM is poorly absorbed and likely serves as nutrients for proinflammatory commensal bacteria in the colon. In contrast, LFM is well absorbed in the proximal gut and is not available to colonic bacteria. Thus, it is conceivable that LFM modulates visceral nociception by altering gut microbiota and preventing intestinal inflammation.

It should be noted that rodent and human microbiota may differ significantly. Hence, caution should be exercised in extrapolating our data to explain human disease. However, it appears that foods differing in FODMAP composition induce some similar changes in colonic microbial compositions in rats and humans (27). In a human clinical study, we observed elevated fecal LPS levels among IBS-D patients compared with HS (Figure 9A). Four weeks of treatment with LFM diet may not only reduce abdominal pain and bloating but also normalize the fecal LPS to a level similar to that of HS. Similar to our findings in rats, we observed that intracolonic administration of fecal supernatant from IBS-D patients caused visceral hypersensitivity (Figure 9, B and C). Administration of LPS inhibitor, LPS-RS, prevented the development of visceral hypersensitivity caused by fecal supernatant of IBS-D patients (Figure 9, B and C). Following 4 weeks of LFM treatment, the fecal supernatant from the same 6 IBS-D patients did not evoke an abnormal increase in visceral pain in response to CRD (Figure 9, B and C). These findings confirm our observation in rats that FODMAP diet modulates visceral nociception by LPS-mediated intestinal barrier dysfunction and gut inflammation.

In conclusion, we have shown that an HFM diet causes an increase in fecal LPS, likely from gut dysbiosis. This induces mucosal inflammation, impairs permeability, and contributes to the development of visceral hypersensitivity. In contrast, an LFM diet reduces fecal LPS by modulating gut microbial composition. This decreases mucosal inflammation, improves gut barrier function, and prevents stress-induced visceral hyperalgesia. Similar observations were made in IBS-D patients who show an elevated fecal LPS level that was normalized following 4 weeks of treatment with LFM. Fecal supernatant from IBS-D patients receiving LFM diet also failed to evoke visceral hypersensitivity in naive rats. These observations may explain how an LFM diet benefits IBS patients.

### Methods

#### Study approval. All experimental procedures were performed in accordance with NIH guidelines and approved by the University Committee on Use and Care of Animals at the University of Michigan. The protocol for human colonoid culture was approved by the Institutional Review Board of the University of Michigan Hospital with written informed consent.

#### Animals, diets, and experimental procedures. Adult male Wistar rats (200–220 g) were housed 3 per cage in a controlled environment (12-hour daylight cycle, lights off at 18:00) with free access to food and water. The composition of the HFM diet was based on a human clinical study (2): 10% w/w FODMAPs, comprising 3.5% w/w fructose, 3.5% w/w lactose, and 3% w/w oligofructose (D12020103, Research Diets). Rats were randomized into groups, and for 14 days were fed HFM or RC (2.1% FODMAPs). Rats were treated with oral gavage of rifaximin (150 mg/kg) (16) or water twice daily, 8 hours apart, for 14 days. In a separate study, rats were fed RC or an LFM diet (0% FODMAPs) (D12020102, Research Diets) for 14 days, and then exposed to daily WAS for 10 days or daily RS for 10 days. Each gram of HFM and LFM contains 3.89 and 3.96 kcal, respectively, with 6.7% and 6.9% of calories, respectively, provided by fat, 61.4% and 63.8% of calories provided by carbohydrate, and 19.5% and 19.9% of calories provided by protein.

#### Repeated RS. Repeated RS was conducted as described previously (16). Each rat was submitted to a 120-minute restraint period in a plastic rat restrainer (diameter 6.5 cm and length 15 cm), daily, for 10 consecutive days. This restraint procedure minimized the space around...
Sequences were classified using the Ribosomal Database Project (RDP) 16S rRNA gene training set (version 9) using a naive Bayesian approach with an 80% confidence threshold. Sequences that did not classify or were identified as Eukaryota, Mitochondria, Chloroplasts, Archaea, or Unknown were removed. We used the cluster.split command in mothur to cluster sequences into operational taxonomic units (OTUs) at a cutoff equal to 0.03. OTU sequence data were converted to relative abundance ± SEM.

Quantitative or semiquantitative RT-PCR for inflammatory cytokines. Total RNA was extracted from proximal colon tissue samples using TRIzol reagent (Life Technologies), according to the manufacturer’s instructions. cDNA was synthesized using iScript cDNA synthesis kit (Bio-Rad Laboratories). Quantitative PCR (qPCR) for inflammatory cytokines and glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was performed with a CFX Connect Real-Time PCR Detection System (Bio-Rad Laboratories) using SYBR Green detection.

Primers used for qPCR, GAPDH, IL-1β, IL-6, IL-10, TNF-α, and IFN-γ were obtained from Qiagen. Primer sequences for IL-17 are as follows: forward IL-17, 5′-ACAGTGAAGGCAGCGGTACT-3′; reverse IL-17, 5′-GCTCAGAGTCCAGGGTGAAG-3′.

The PCR conditions were as follows: one cycle at 95°C for 10 minutes, followed by 40 two-temperature cycles at 95°C for 15 seconds and 60°C for 60 seconds. PCR amplifications were performed in a total volume of 25 μl, containing iQSYBR Green supermix (Bio-Rad Laboratories). Cytokine transcript levels were normalized to that of GAPDH, and relative gene expression was expressed as the fold change (2^−ΔΔCt) relative to expression in the control samples.
Western blot analysis. Proteins were extracted from the proximal colon tissues and analyzed on Ready Gel Tris-HCl (Bio-Rad Laboratories). The tissues were homogenized in RIPA buffer (1% IGEPAK, 0.5% sodium deoxycholate, and 0.1% SDS in Tris-buffered saline solution [pH 7.4]), supplemented with protease inhibitor cocktail (Sigma-Aldrich). The homogenate was centrifuged at 14,000 g for 10 minutes. Equal amounts of protein (30 μg) were separated by 4–20% Ready Gel Tris-HCl gels (Bio-Rad Laboratories), transferred to polyvinylidene difluoride membranes, and blocked with StartingBlockT20 blocking buffer (Thermo Fisher Scientific) for 60 minutes at room temperature. Membranes were incubated with rabbit anti-ZO-1 antibody (61-7300, Life Technologies) and rabbit anti-OCLN antibody (71-1500, Life Technologies) at 1:400 dilution at 4°C overnight, and then washed in Tris-buffered saline for 1 hour. The membranes were then probed with peroxidase-conjugated goat anti-rabbit IgG at 1:8,000 dilution for 1 hour at room temperature, and the bands were visualized by electrochemiluminescence (ECL, Thermo Fisher Scientific). Signals were quantified using ImageJ (NIH) and normalized to controls.

Histology and immunohistochemistry. Segments of the proximal colon were fixed in 10% formalin. For morphologic analysis, colon segments were embedded in paraffin, sectioned at 5-μm thickness, and subsequently stained with H&E. Neutrophils, mononuclear cells, and eosinophils were identified in H&E-stained sections; mast cells were identified in toluidine blue-stained sections. Cells were counted at a magnification of ×400 in 8 different areas above the muscularis mucosae of each section using a micrometer grid and expressed as the number of cells/mm².

Serum and fecal LPS levels. LPS levels were measured with a quantitative chromogenic limulus amoebocyte lysate (LAL) QCL-1000 test kit (LONZA), following the manufacturer’s protocols. Serum and fecal samples were prepared in pyrogen-free water provided with the kit. Dilute serum samples were deactivated at 75°C for 10 minutes in a water bath. Samples were further incubated for 10 minutes with 50 μl of LAL reagent at 37°C, followed by the addition of 100 μl of LAL chromogenic substrate for 6 minutes. The reaction was terminated by adding 100 mg/ml of SDS, and the yellow color that developed due to cleavage of the substrate was measured spectrophotometrically at 405 nm.

TEER. The intestinal barrier function was assessed by measuring the TEER of ex vivo tissues as reported previously (41, 42). TEER reflects paracellular resistance imparted by tight junctions and the lateral paracellular space and is a sensitive measure of barrier integrity (43).

Intestinal tissue from the proximal colon was isolated. Intestinal segments were opened along the mesenteric border, washed in phosphate-buffered saline (PBS), and cut into 5 × 7 mm pieces. Tissues were washed twice in sterilized PBS and transferred to Petri dishes containing DMEM culture medium. After a 30-minute incubation at 37°C and pH stabilization, the TEER was measured using the micro-Snapwelling system with an Endohm SNAP electrode attached to an EVOM2 epi-pH stabilization, the TEER was measured using the micro-Snapwell. After a 30-minute incubation at 37°C and pH stabilization, the TEER was measured using the micro-Snapwell. The tissue was then washed twice in sterilized PBS and transferred to Petri dishes containing DMEM culture medium. After a 30-minute incubation at 37°C and pH stabilization, the TEER was measured using the micro-Snapwell. The tissue was then washed twice in sterilized PBS and transferred to Petri dishes containing DMEM culture medium. After a 30-minute incubation at 37°C and pH stabilization, the TEER was measured using the micro-Snapwell.
Intraluminal cytokine measurements. Human colonoids were incubated with either a mixture of cytokines (10 ng/mL TNF-α, 20 ng/mL IL-1β, 20 ng/mL IFN-γ, and 20 ng/mL IL-18), control medium (optiMEM), or 10 μg/mL LPS. Cytokine concentrations were measured using a bioluminescent immunoassay kit (Invitrogen).

Intracolon siRNA administration. Intracolon application of siRNA was performed as described previously (48, 49). Rats were fasted overnight and then placed in a plastic holder for 30 minutes to empty the bowel contents. Lipofectamine 2000 (Invitrogen) was diluted with OptiMEM (Invitrogen) according to the manufacturer’s instructions and incubated for 5 minutes at room temperature. siRNA targeting TLR4 (Thermo Fisher Scientific) was dissolved in OptiMEM. These 2 solutions were then mixed at a 1:1 ratio and incubated at room temperature for 20 minutes. The combined Liposomal siRNA (300 μM) containing 10 nmol siRNA was then immediately injected into the rat colon at day 0 and day 1 using a flexible plastic tube (18-gauge, 3 inches, Instech Laboratories) inserted into the distal colon 3 inches from the anus. Experiments were perfused 3 days after the first siRNA injection.

Human colonoid culture. The protocol for colonic crypt isolation and culture was a modification of previously described protocols (50, 51). Colonic crypts were obtained from colonoscopy biopsy samples at the University of Michigan Hospitals. The study was approved by the Institutional Review Board of the University of Michigan Hospital with written informed consent. Patient biopsy samples were collected from 5 normal subjects. Colon biopsies were washed with PBS containing penicillin-streptomycin (Pen/Strep, 1× Invitrogen), gentamicin (50 μg/mL), Life Technologies), Normocin (100 g/mL, InvivoGem), and amphotericin B (2.5 μg/mL, Sigma-Aldrich) to control contamination, with thiazovivin (2.5 μM, Stemgent), a potent inhibitor of Rho-associated coiled-coil-containing protein kinase (ROCK). After 3 washes over 30 minutes, tissue was incubated in nonenzymatic-digestion PBS containing 8 mM EDTA (Lonza) with DTT (Sigma) for 5 minutes to remove mucus and in 8 mM EDTA for 5–10 minutes for further digestion. All digestion-buffer PBS contains Pen/Strep (1×) and Normocin (100 μg/mL). Colonoids were released by shaking PBS containing Pen/Strep (1×) for 1–2 minutes. Isolated crypts were centrifuged at 500 g for 1 minute. Colonic crypts were collected and plated in 24-well plates with growth factor–reduced Matrigel (Corning). The Matrigel was polymerized for 10 minutes at 37°C. R-spondin (50 ng/mL) and Wnt3a-conditioned advanced DMEM/F12 culture medium were added to each well. The culture medium contained 1× GlutaMax, HEPES, B27, insulin-transferrin-selenium (ITS), Pen/Strep, N-acetylcysteine (1 mM), nicotinamide (5 mM), as well as growth factor combinations, including murine EGF (100 ng/mL), nogo (100 ng/mL), A8301 (0.5 μM), SB202190 (30 μM), Chiron (10 μM), PGE2 (10 μM), thiazovivin (2.5 μM), and diltated anoikis inhibitor. After several passages, colonoids were cultured with fecal supernatant. After 24 hours, colonoids were harvested, and RNA was extracted for qPCR.

Colonoid microinjection. Microinjections were performed on human colonoids with a mixture of fecal supernatant, LPS, or LPS-RS with FITC–dextran. Thin-wall glass pipettes (TW100F-4; World Precision Instruments) were pulled using a Narishige PN-30 micropipette puller. The glass pipette tips were cut open and filled with a mixture containing 27 μL of fecal supernatant, 3 μL of 4-kDa FITC–dextran (10 mM, Sigma-Aldrich), and LPS (10 μg/mL) or LPS-RS (100 μg/mL) using Eppendorf Microloader pipette tips. Microinjections were performed with a XenoWorks BRI analog microinjector (Sutter Instrument). After injection, colonoid spheres were selected for 24-hour live-cell imaging. To determine the epithelial barrier function of the colonoid spheres after application of the supernatant, images of FITC–dextran in the colonoid spheres were captured every 30 minutes using the DeltaVision epifluorescence imaging system (GE Healthcare Bio-Sciences) at ×4 magnification and the pixel intensity measured. Colonoid spheres (n = 6–10) were selected for 24-hour live-cell imaging. Images were recorded by SoftWoRx imaging software (Applied Precision, GE Healthcare Bio-Sciences) for off-line analysis. The disruption of barrier integrity was visualized by the loss of FITC–dextran in the lumen of each colonoid sphere compared with the starting point (52).

Statistics. EMR amplitude is represented by the area under the curve. The effects of stress and/or dietary treatment on VMR to CRD were analyzed by comparing poststress and baseline values at each digestion pressure, using 2-way repeated-measures ANOVA. To examine data for other studies, differences between groups were compared by 2-tailed Student’s t test or ANOVA for comparisons between 2 groups or more than 2 groups, respectively. Results are expressed as the mean ± SEM. P less than 0.05 was considered statistically significant.

Author contributions. CO and SYZ conceived and designed the study, SYZ, MG, XW, GZ, HZ, YL, BY, and PL acquired the data. SYZ and MG analyzed and interpreted the data. SYZ, MG, and CO wrote the manuscript. CO obtained funding.

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